

# The First Few X cases and contacts (FFX) investigation template protocol for respiratory pathogens with pandemic potential







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# **Abbreviations**

**CONSISE** Consortium for the Standardization of Influenza Seroepidemiology

**COVID-19** Coronavirus disease 2019

**FFP2** Filtering Face Piece

**FFX** First Few X cases and contacts

**GIP** Global Influenza Programme

**GOARN** Global Outbreak Alert and Response Network

**HLIP** High-level Implementation Plan (Pandemic Influenza Preparedness Framework)

**IHR** International Health Regulations

**ILI** Influenza-Like Illness

**IQR** Interquartile Range

**MERS-CoV** Middle Eastern Respiratory Syndrome coronavirus

**PCR** Polymerase Chain Reaction

**PIP** Pandemic Influenza Preparedness

**PPE** Personal Protective Equipment

**R**<sub>0</sub> Basic reproductive number

**R**<sub>eff</sub> Effective reproductive number

**RNA** Ribonucleic acid

**SAR** Secondary Attack Rate

**SARI** Severe Acute Respiratory Infection

**SARS-CoV-2** Severe acute respiratory syndrome coronavirus 2

**SCAR** Secondary Clinical Attack Rate

**SIR** Secondary Infection Rate

**WHE** Health Emergencies Programme

**WHO** World Health Organization

# **Summary**

This document sets out the methods to guide data collection and the public health investigation for the **comprehensive assessment of human case(s) of a novel or re-emerging respiratory pathogen with pandemic potential and their close contacts.**For the purposes of this protocol, the conceptual respiratory pathogen in question will be referred to as pathogen X, which causes Disease X. This protocol provides guidelines for conducting a First Few X cases and contacts (FFX) investigation for pathogen X in [Country Y], as part of the Respiratory Investigations and Studies, Unity Studies initiative.

The detection and spread of a novel or re-emerging pathogen with pandemic potential is accompanied by uncertainty over the key epidemiological, clinical and virological (if applicable) characteristics of the novel pathogen, particularly its ability to spread in the human population and its virulence (case-severity). As a result, it is essential to understand the epidemiological, clinical and virological (if applicable) characteristics of the First Few X cases (FFX) of this novel or re-emerging pathogen and their close contacts, in order to inform targeted guidance and measures for the public health response.

The World Health Organization (WHO), in collaboration with technical partners, has developed a series of enhanced surveillance protocols that are harmonized to help provide detailed insight into the epidemiological characteristics of respiratory pathogens with pandemic potential. They build on previous protocols developed by the global Consortium for the Standardization of Influenza Seroepidemiology (CONSISE) (1), and WHO's Influenza Pandemic Special Investigations and Studies, as well as those developed by WHO as part of the Unity Studies for COVID-19 (2) and for Middle East respiratory syndrome coronavirus (MERS-CoV) (3).

All WHO Respiratory Investigations and Studies protocols are available on the WHO website. CLICK TO VIEW

**Comments** for the user's consideration are provided in purple text throughout the document, as there may be a need to modify methods slightly considering the local context in which the investigation will be carried out.

Full title of study	The First Few X cases and contacts (FFX) comprehensive investigation template protocol for respiratory pathogens with pandemic potential [Country Y]							
Population	The First Few X cases of pathogen X and their close contacts							
Potential output and analysis	Transmission dynamics, severity and clinical spectrum in the population of initial cases of <b>pathogen X</b> through estimates of, primarily:							
	<ul> <li>the secondary infection rate (SIR) and secondary clinical attack rate (SCAR) overall, and by key factors such as setting, age and sex;</li> <li>the clinical presentation and course of associated disease;</li> <li>the symptomatic and asymptomatic proportions of cases, and;</li> <li>preliminary case- (i.e. disease) hospitalization and fatality ratios, and infection-hospitalization and fatality ratios.</li> </ul>							
	And secondarily:							
	<ul> <li>the serial interval;</li> <li>duration of viral shedding (if repeated virological samples are taken at a high frequency where adequate resources are available);</li> <li>possible routes of transmission including possible animal-human transmission, and;</li> <li>risk and/or protective factors for transmission or severe disease.</li> </ul>							
	Advanced related objectives:							
	<ul> <li>the basic reproduction number (R<sub>o</sub>);</li> <li>the effective reproduction number (R<sub>eff</sub>);</li> <li>the incubation period, and;</li> <li>the generation time.</li> </ul>							
Study design	Prospective case-ascertained investigation of all identified close contacts of pathogen X index cases.							
Timing of the	To be initiated in the first days after the confirmation of a case of pathogen X in [Country Y].							
investigation	FFX is the primary protocol to be initiated in the case of a pathogen X outbreak/cluster, upon identification of the initial cases of pathogen X in [Country Y] in the early epidemic/pandemic phases.							
Duration	Recruitment and follow-up of index cases and all close contacts for 28 days from recruitment.							
Minimum data and specimens to be obtained from participants	Epidemiological, clinical, virological (if applicable i.e. if pathogen X is a virus) and serological data will be collected from each participant at multiple times during the investigation – including surveys at baseline and day 28, symptom diaries from days 2-28 and specimen collection.							

# 1 Background and objectives



## 1.1 Background to Pandemic Investigations and Studies

Following a review of the global response to the last influenza pandemic (2009 influenza pandemic H1N1) (4), the global Consortium for the Standardization of Influenza Seroepidemiology (CONSISE) (1), and World Health Organization's (WHO) Influenza Pandemic Special Investigations and Studies initiative were established to develop a suite of standardized early investigation protocols, supported by the Global Influenza Programme (5) and the Pandemic Influenza Preparedness (PIP) Framework (6). Standardized protocols were also implemented following the emergence of Middle East respiratory syndrome coronavirus (MERS-CoV) in 2012 (3), and Zika virus in 2016 (7).

In January 2020, the novel coronavirus, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), responsible for the coronavirus disease 2019 (COVID-19) was declared a public health emergency of international concern by the WHO. This required rapid implementation of early investigations to inform appropriate national and global public health actions. Adaptation of existing protocols was initiated from the first weeks of the detection of the novel coronavirus and further developed under the WHO initiative, Unity Studies (2). The Unity Studies protocols standardized methods and facilitated rapid generation of local data for public health action and comparison of key epidemiological parameters across regions and globally (2, 8–10).

Based on the learning of the COVID-19 response, a series of standardized template protocols have been developed for both disease specific investigations such as influenza and for any novel respiratory pathogen of pandemic potential for the implementation of standardized and quality investigations and studies to ensure readiness in advance of a future pandemic. These have been developed as part of the WHO Investigations and Studies, Unity Studies initiative. CLICK TO VIEW The initiative aims to provide an 'at the ready' international framework for preparedness and response to future pandemics, providing a suite of enhanced surveillance and investigational activities that are harmonized to help provide detailed insight into the epidemiological characteristics of emerging or re-emerging respiratory pathogens of pandemic potential. They are included in both the Global Influenza Strategy (11) (pillar 6: "Number of sites (or geographical coverage) primed to conduct at least one of the WHO Pandemic Influenza Special Investigations in case of a pandemic with target for 2023: "At least 2 operationally ready sites in each WHO region (so 12 sites)") and the third version of the High-Level Implementation Plan (HLIP III) CLICK TO VIEW for pandemic preparedness (PIP) (12) for the 2024-2030 period under Output Indicator 2.11 "Number of sites participating in the WHO Investigations and studies network (Unity Studies)".

Investigations and Studies are also included in the Mosaic Respiratory Surveillance Framework (13). The Mosaic framework presents respiratory virus surveillance systems into a collaborative context where they are each focused on the objectives to which they are best suited. As it is impossible to address the many complex needs of respiratory virus surveillance with a single system, multiple fit-for-purpose surveillance approaches and complementary investigations must fit together as tiles in a "mosaic". Only together will these

approaches provide a complete picture of respiratory viruses and the impact of associated illnesses and interventions at the country level (13). Discrete studies and early investigations such as the Unity Studies can address certain public health objectives that are not efficiently met by existing systems such as rapidly assess transmissibility, estimate population susceptibility/immunity and infection severity, aid identification of population groups in need to target interventions, and estimate burden of disease and vaccine effectiveness (13).

The detection and spread of respiratory pathogens with pandemic potential, or disease X, are accompanied by scientific uncertainty relating to their epidemiological and serologic characteristics, transmissibility (i.e., ability to spread in a population), and virulence (i.e. i.e., severity). Looking ahead, leveraging smart surveillance approaches through protocol implementation and standardization will be key to prepare for future potential panrespiratory pathogen threats (14). These types of enhanced surveillance investigations can be used to inform public health responses to respiratory pathogens of pandemic potential. Pathogen X might be a novel pathogen (e.g., SARS-CoV-2 in late 2019) or a re-emerging existing pathogen (e.g., novel strains of influenza).

# 1.2 Introduction to FFX studies and their approach

As with many novel respiratory pathogens, key epidemiological, clinical and virological (if applicable i.e. if pathogen X is a virus) parameters of the pathogen and the outbreak dynamics are unknown at the beginning. This is the situation for pathogen X, first detected in [Country Y] in [mm/yyyy].

At this stage, key questions exist around the transmissibility and infection-severity of pathogen X. As a result, it is essential to understand the epidemiological, clinical and virological characteristics of the First Few X cases (FFX) of pathogen X and their close contacts, in order to inform targeted guidance and measures for the [Country Y] public health response. FFX investigations are intended to include the first confirmed cases of a novel respiratory pathogen detected in [Country Y] although the exact number of confirmed cases included will depend on the nature of the pathogen as well as country resources. See further details in **Section 3.1.** 

This FFX investigation will be conducted across several pre-selected countries or sites with geographic and demographic diversity. These sites will be part of a network of pre-determined and capacity built sites (WHO Unity Studies sites). However, any country can use and implement the FFX investigation protocol.

By using a standardized protocol such as the one described here, epidemiological exposure data and biological samples can be systematically collected and shared rapidly in a format that can be easily aggregated, tabulated and analyzed across many different settings globally. This will facilitate timely estimates of the infection-severity and transmissibility of pathogen X infection, as well as informing public health responses and policy decisions.

Each country may need to tailor some aspects of this protocol to align with local public health, laboratory and clinical systems, according to their country capacity and availability of resources, as well as the cultural appropriateness of the protocol. The protocol should also align with country plans for case and contact clinical and public health management, including infection prevention and control measures for both health workers and the cases and contacts.



#### **Toolkit item**

The timely and standardized implementation of this protocol should be supported by a toolkit developed by WHO and implementing partners. The toolkit will comprise components to support different elements of the protocol. These will be highlighted throughout the protocol and discussed in Section 5. CLICK TO VIEW TOOlkit components will be available on the WHO website.

Toolkit items to support this section may include:

- · Checklist for Unity Studies alignment
- Terms of Reference for Unity Studies network sites

See additional 'Toolkit item' boxes throughout this document.

This **FFX protocol** outlines the process for rapid data collection from early cases of pathogen X and all their close contacts. FFX studies will provide critical early insight into key epidemiological characteristics such as the transmissibility and severity of pathogen X infection. This protocol **may be the first investigation to be conducted**.

of household and health worker contacts, and then expand to other close contacts if resources allow. However, this may be better addressed through other Respiratory Investigations and Studies of pathogen X – namely the household transmission investigation and closed setting protocols. These studies can be undertaken simultaneously or subsequently, to collect further information relating to pathogen X infection, depending on the availability of resources and capacity. These studies include prospective investigations of transmission of pathogen X in households and in closed environments, such as in schools, health facilities/hospitals¹, army barracks, nursing homes, and planes.

These investigations can:

- o Reinforce findings from FFX and other investigations, and;
- o Produce setting-specific estimates of key transmissibility and severity parameters including the effect of interventions in reducing the risk of infection; the risk of secondary infection, and; the asymptomatic fraction.

<sup>1</sup> Health workers at risk of infection include: ambulance staff, reception staff, health assistants, nurses, doctors, laboratory workers and cleaners.

The table in **Appendix A CLICK TO VIEW** summarizes the different features and complementarity of the three protocols. All protocols are available on the **WHO website**.

# 1.3 Objectives

The overall aim of this protocol is to gain an early understanding of key clinical, epidemiological and virological (if applicable) characteristics of the first cases of pathogen X infection detected in [Country Y]. This information will be used to refine/ update recommendations for surveillance and public health guidance (for example, case definitions); to characterize the key epidemiological transmission features of the pathogen; to help understand the geographic spread, severity and impact on the community; and to inform operational models for the implementation of countermeasures such as non-pharmaceutical interventions (15) (for example, case isolation, contact tracing, etc) and medical interventions, if possible.

**Comment:** 

Further details to be added here according to the specific context of the epidemic/pandemic.

**Comment:** 

FFX studies can also be run during later stages of an outbreak or with other objectives relating to ongoing characterization of epidemiology. These study design adaptations may require different data and specimen collection, as outlined in **Section 2.5** 

click to view and 2.6. click to view

The ability of an FFX investigation to answer each transmissibility or severity objective below will ultimately depend on the type and frequency of data and/or specimen collection. The rationale for specimen sampling is provided in Section 2.6. CLICK TO VIEW

# 1

#### **Primary objectives**

The **primary objectives** of the FFX investigation among cases and close contacts are to provide descriptions or estimates of <u>transmissibility</u> and <u>severity</u>:



#### **Transmissibility**



#### Severity

- Secondary infection rate (SIR)<sup>2</sup> of pathogen X infection overall, and by key factors such as setting, age and sex;
- 2. Secondary clinical attack rate (SCAR) as a proxy measure of pathogen X infection among close contacts, overall, and by key factors such as setting, age and sex;
- 1. Clinical presentation of pathogen X infection and course of associated disease;
- 2. Symptomatic and asymptomatic proportions of pathogen X cases, and;
- 3. Preliminary case- (i.e., disease) and infection-hospitalization and fatality ratios.

### 2

#### **Secondary objectives**

The **secondary objectives** are to provide data to support the estimation of further characteristics of the <u>transmissibility</u> and <u>severity of pathogen X</u>:



#### **Transmissibility**



#### Severity

- 1. Serial interval of pathogen X;
- 2. Duration of viral shedding (if repeated virological samples are taken at a high frequency where adequate resources are available);
- 3. Possible routes of transmission including possible animal-human transmission, and;
- 1. Risk and/or protective factors for transmission or severe disease.



#### **Advanced related objectives**

The **advanced related objectives**, to be addressed with the inclusion of modelling or genomic analysis, enable further characterisation of the <u>transmissibility of pathogen X</u>:



#### **Transmissibility**

- 1. Basic reproduction number  $(R_{o})$  of pathogen X;
- 2. Effective reproduction number  $(R_{off})$  of pathogen X;
- 3. Incubation period of pathogen X, and;
- 4. Generation interval of pathogen X.

<sup>2</sup> Alternative terminology for this parameter is the "secondary infection risk". It represents an overall risk of infection among close contacts for a defined time period. "Secondary infection rate" is used here as this term is widely used and recognised throughout the literature.

# The following glossary provides definitions for key epidemiological terms used in this protocol.

- The **secondary infection rate (SIR)**<sup>2</sup> is a measure of the frequency of new **infections** of pathogen X among contacts of confirmed primary cases in a defined period of time, as determined by a positive pathogen X laboratory result. *In other words, it is the rate of contacts being infected, assessed through polymerase chain reaction (PCR), virus culture, or serological assays on paired samples.*
- The **secondary clinical attack rate (SCAR)** is a measure of the frequency of new symptomatic persons among contacts in a defined period of time<sup>3</sup>. In other words, it is a proxy measure of the SIR, representing the rate of clinical manifestation of the infection in contacts it is dependent on the clinical criteria being used in case definitions.
- The **asymptomatic proportion of cases** is a measure of the frequency of asymptomatic infections of pathogen X among all confirmed cases in a defined period of time. *In other words, it represents the proportion of laboratory confirmed cases who do not display symptoms of disease an individual's symptom status is dependent on the clinical criteria being used in case-definitions.*
- The **infection-hospitalization ratio**<sup>4</sup> is defined as the proportion of persons with a laboratory confirmed pathogen X infection who are admitted to hospital for clinical management or treatment.
- The **infection-fatality ratio**<sup>4</sup> is defined as the proportion of persons with a laboratory confirmed pathogen X infection who die as a direct or indirect consequence of their infection.
- The **serial interval** is defined as the period of time from the onset of symptoms in the primary case to the onset of symptoms in a secondary case.
- The **basic reproduction number**  $R_o$  is defined as the number of infections produced, on average, by an infected individual in the early stages of the epidemic, when virtually all contacts are susceptible. Note that this assumes that there is little-to-no population-level immunity to pathogen X.
- Once interventions are put in place or the number of susceptible individuals declines, the transmission potential of the disease at a given time (t) is measured in terms of the effective reproduction number R<sub>eff</sub>

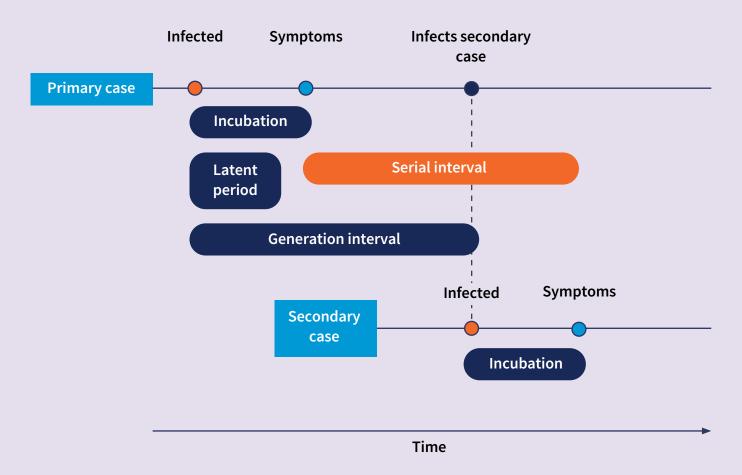
<sup>3</sup> This can be determined by a positive pathogen X result along with presence of symptoms, or by only the presence of symptoms, however relying on symptoms alone could result in incorrect classification of contacts as the cause of symptoms may be from other respiratory pathogens and not pathogen X.

<sup>4</sup> If laboratory confirmation is not available for all contacts, there may be undetected infections. In this instance, only **case-hospitalization and case-fatality** ratios can be calculated.

- The **incubation period** is defined as the period of time between an exposure resulting in pathogen X infection and the onset of the first clinical symptoms of the disease.
- The **generation interval** is defined as the period of time from infection in a primary case to infection of a secondary case.
- The **latent period** is defined as the period of time between when an individual is infected by pathogen X and when they become infectious.
- The **duration of viral shedding** is the time for which pathogen X is shed, regardless of clinical symptoms.

The relationship of time periods related to a primary and a secondary case and the main epidemiological parameters can be summarized as below (Figure 1).

**Figure 1:** Relationship of time periods related to a primary and a secondary case and the main epidemiological parameters



### 1.4 Overview of methodology

Section 2 of this protocol will describe in detail the methodology for this investigation including the study design, start and duration of the investigation, case and contact definitions, data collection, data management, specimen collection and transport, laboratory testing and ethical considerations.

Coordination of investigations and sharing of information in real-time will be needed at both country and global levels. Epidemiologists, modelers, virologists, statisticians, clinicians and public health experts will all assist in developing early estimates of key clinical, epidemiological and virological parameters of pathogen X. Table 1 shows the roles and responsibilities involved for [Country Y].

**Table 1:** Coordination matrix of roles and responsibilities in [Country Y]

What?	Who?
Overall coordination of the investigation	[Cite institution/body/person(s)]
Case detection and investigation	[Cite institution/body/person(s)]
Contact identification and follow-up	[Cite institution/body/person(s)]
Analysis of data	[Cite institution/body/person(s)]
Data management	[Cite institution/body/person(s)]
IT management	[Cite institution/body/person(s)]
[add more roles, as per country context]	[Cite institution/body/person(s)]

The FFX system will be maintained centrally by [cite institution/body/person(s)]. Centralized coordination will require development of a "command and control" plan, to allow for prioritization of investigations.

It is noted that undertaking this kind of enhanced investigation, will require resources and implementation plans need to be developed in advance.



#### **Toolkit item**

Toolkit items to support this section may include:

- Funding guidance
- How to access support
- Project management/budgeting guidance

Toolkit components will be available on the WHO website. CLICK TO VIEW

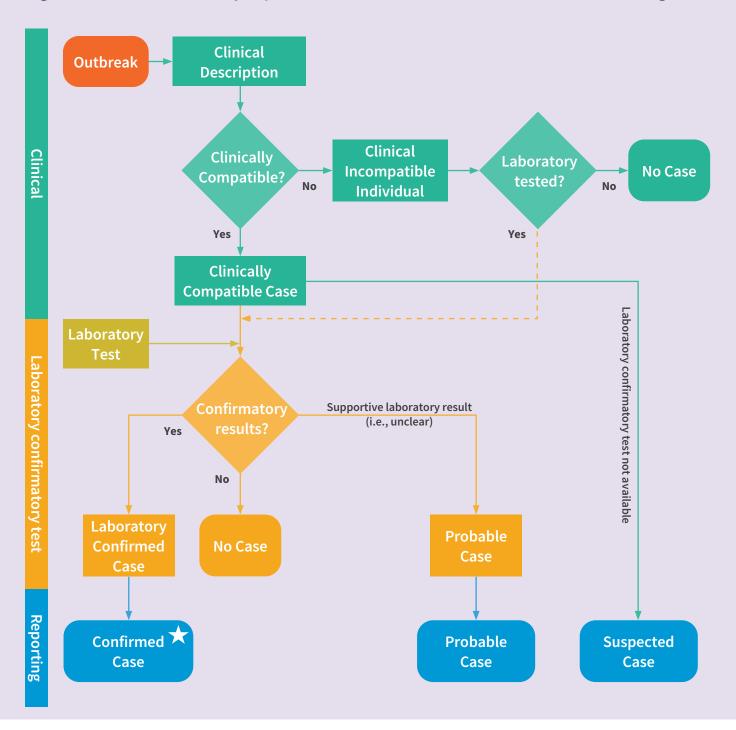
Methods

# 2.1 Definitions used in the FFX protocol

#### 2.1.1 Case definitions

Case definitions are necessary to standardise the identification of cases. A case definition should include when, where, and who is affected, **and outline all criteria determining how an individual is identified as a case**.

Figure 2: Clinical and laboratory steps for the identification of a confirmed case in an FFX investigation.



For pathogen X, cases must be reported as *Confirmed, Probable or Suspected* cases based on a series of epidemiological, clinical and laboratory criteria (**Figure 2** CLICK TO VIEW ) to ensure data specificity and comparability.

A summary of relevant definitions that will be used to form the case definitions (16) for pathogen X include:

- 1) Clinical Description: A description of the illness signs, symptoms (e.g., cough, sore throat, fever), severity, and individual characteristics where relevant (e.g., international travel history) associated with the outbreak;
- 2) Clinically compatible case: An individual experiencing a clinical syndrome compatible with the clinical description in (1);
- 3) Laboratory confirmed case: a case that is confirmed by one or more of the laboratory methods listed in the case definition under laboratory criteria for diagnosis, irrespective of clinical criteria in (2);
- 4) Supportive or presumptive laboratory result: meets the clinical criteria in (2), yet does not fully meet the criteria for laboratory confirmation due to unclear results from laboratory testing

#### **Comment:**

In the early stages of an outbreak of pathogen X, the appropriate identification and enrolment of index cases contributes to the internal validity of the FFX study (17).

The case definitions for pathogen X reporting will be published on the WHO website when available.

For the purpose of this protocol and with these definitions in mind, **the generic interim case definitions for pathogen X are proposed in Box 1.** These definitions will be subject to change as more information and additional diagnostics become available. For instance, the case definition may be adapted to incorporate point of care or rapid antigen tests with proven sufficient sensitivity and specificity as they become available.

#### **Comment:**

An index case is the first case of pathogen X reported or identified within a cluster. It is the identification of the index case that leads to recruitment into the study. The definition of an index case is explored further in **Section 2.1.3**.



#### Box 1. Case definitions in the FFX protocol

#### **Confirmed case:**

A person with laboratory confirmation of pathogen X infection, irrespective of meeting the clinical description (i.e., irrespective of clinical signs and symptoms). Laboratory confirmation includes receiving a positive result from polymerase chain reaction (PCR), virus isolation or from other validated laboratory methods.

#### **Probable case:**

A person meeting the clinical description and has a supportive or presumptive laboratory result

#### **Suspected case:**

A person meeting the clinical description, with no laboratory confirmation or testing results.

#### **Comment:**

The enrolment of confirmed index cases into FFX investigations must be prioritised where possible. In a scenario where a laboratory confirmatory test is not yet available, it may be feasible to recruit suspected index cases (as defined above and according to the WHO guidance for Pathogen X) as the starting point for an FFX investigation, however they would need to be recorded as non-laboratory confirmed. The limitations of using non-laboratory confirmed cases must be noted and accounted for in the analysis. Where feasible, consider if samples from suspected cases could be collected and stored for future testing when a confirmatory test becomes available. For these cases, the outcome of laboratory diagnosis should be recorded and reflected in data analysis as soon as it becomes available.

#### 2.1.2 Contact definitions

**Contacts** are defined as all individuals who are associated with some sphere of activity of the case and may have similar, or other, exposures as the case. Contacts can include household members, social or health workers, other family contacts, visitors, neighbours, colleagues and co-workers, teachers, classmates, and members of a social group.

As with case definitions, contact definitions for pathogen X will be available on the WHO website. These definitions will be subject to change as more information becomes available about pathogen X.

FFX investigations focus on **close contacts**, which are a subset of contacts who meet a minimum level of interaction with the case.

The definition of close contact should consider:

- Plausible modes of transmission (e.g., droplets, aerosols, fomites);
- Duration of contact;
- Setting of contact (i.e., indoor or outdoor);
- Infectiousness of pathogen X relative to symptom onset;
- The period of time from symptom onset in an index case (see 2.1.3 CLICK TO VIEW ) to testing, notification and recruitment into the FFX investigation.

For the purpose of this protocol, interim and generic **close contact** definitions and classification for pathogen X are proposed in **Box 2**.

#### **Comment:**

Implementing countries may wish to consider more detailed definitions of close contacts if appropriate.



#### Box 2: Close contact definitions in the FFX protocol

#### **Close contact:**

Any person who had contact (within 2 meters and for more than 5 minutes):

- with a symptomatic case during their **symptomatic** period, 2 days before symptom onset, and in the 10 days after the onset of symptoms, or;
- with a confirmed **asymptomatic** case, including 2 days before and the 10 days after the date on which the sample was taken which led to confirmation.

**Comment:** 

The contact does not have to be direct physical contact

#### **Further classification of close contact** (for use in contact questionnaires):

- **Health worker contact:** any social or health worker\* who provided direct or indirect personal or clinical care, handling specimens from, or examination of a case of pathogen X, **or** who was within the same indoor space when an aerosol-generating procedure was implemented, **AND** who were not wearing recommended personal protective equipment (PPE)^ at the time or with a possible breach of PPE.
  - \* Health workers at risk of infection with pathogen X include: ambulance staff, reception staff, health assistants, nurses, doctors, laboratory workers and cleaners.
  - ^ Full PPE is defined as correctly fitted high filtration mask (FFP2), gown, gloves and eye protection.
- Household contact: any person who resides or resided in the same household<sup>†</sup> as the pathogen X case\*\*
  - † Household is defined as a group of people (2 or more) living in the same residence. In practice, the technical definition may vary due to social, political and cultural practices (18). Definitions may be (but are not limited to):
    - Two or more people living together in a domestic residence (residential institutions, such as boarding schools, dormitories, hostels, or prisons will be excluded).
    - One possible generic definition of a household is a dwelling or group of dwellings with a shared kitchen or common opening onto a shared household space.
  - \*\* Household contact definitions may vary and implementing countries may wish to consider more detailed definitions if appropriate (see below).

#### A household contact may also be:

- a person who **commonly resides** in the same household as the case
- any person who had **resided** in the same household as the case **for at least one night** during the exposure period (two days before to 10 days after onset of illness in the primary case)

**Comment:** 

It is important that the definitions are consistently applied throughout the investigation and well detailed in any reporting of the investigation for the purposes of comparability between investigations

#### 2.1.3 Classification of FFX participants

During the course of the FFX investigation, transmission events associated with an index case will be observed (or inferred) through laboratory testing and symptom monitoring of close contacts. These observations will help identify the chains of transmission within clusters and allow for final classification of all participants, as described below.



#### **Box 3: Classification of FFX participants**

- **A.** Index case: the first case of pathogen X identified within a cluster according to the case definition (see Box 1). It is the identification of the index case that leads to recruitment into the study\*.
- **B. Primary case:** an individual who has the first evidence of infection/disease within the recruited cluster (See Box 1) i.e., the case with the earliest symptom onset date and/or positive laboratory test within a cluster.
- **C. Co-primary case(s):** of the first cases identified within a cluster, two or more cases identified within the same 24 hour period are considered to be **"co-primary" cases**.

#### **Comment:**

Cases in A), B) and C) must also be classified further as confirmed, probable or suspected cases according to **Box 1**. The relevant clinical and laboratory criteria for pathogen X must be added into the definitions above when available.

- \* In some instances, the index case is also the primary case but this is not always the case. Following investigation, index cases may be classified as a primary, co-primary or secondary case (19).
- **D. Imported case:** an index, primary or co-primary case with a history of travel from an affected area [define "affected area"] in the 14 days before disease onset.
- **E. Secondary cases** are close contacts meeting the:
- 1. Case definition between 24 hours to 10 days after the confirmation or symptom onset of the primary and/or co-primary case(s),

#### OR those demonstrating evidence of;

2. Seroconversion, of which a generalized definition is a 4-fold increase in pathogen X specific antibody titre between paired serum samples, collected at baseline (day 1) and follow-up.

Note: Close contacts meeting either of the above conditions are **not necessarily a secondary case**. Depending on the chains of transmission, an individual meeting these criteria may also be an unrelated case (see F below). It is important to consider the timing of exposure and virological characteristics (if appliable) of pathogen X when classifying close contacts as secondary cases. Testing protocols, symptom information and genomic sequencing data may help to distinguish between secondary cases and unrelated cases.

#### **Comment:**

Cases in E), may also be classified as confirmed, probable or suspected secondary cases according to **Box 1**. The relevant clinical and laboratory criteria for pathogen X must be added into the definitions above.

#### **Comment:**

When laboratory diagnostics are not available for secondary case classification, the limitations of classifying secondary cases according to the suspected or probable secondary case definition must be noted and accounted for in the analysis.

**F. Unrelated case:** includes other cases for the purposes of the FFX investigation such as tertiary cases (those with evidence of being infected by a secondary case) and cases infected from other external sources (i.e., not the primary case)

## 2.2 Study design, duration and population of interest

#### 2.2.1 Study design

This FFX investigation is a prospective case-ascertained investigation of index cases of pathogen X and all identified close contacts, including infants and children. This is distinct from a cohort study in which a group of disease-free households are recruited and then followed over time. Case-ascertained transmission studies are more efficient than cohort studies when interest is in early estimation of the clinical, epidemiological and virological (if applicable) characteristics of an emerging pathogen. This is because the risk of primary or secondary infection in a cohort would be expected to be low during the early stage of the pandemic before widespread community transmission is established.

#### 2.2.2 Timing of the study

This FFX investigation should be established as soon as possible after the identification of the first cases of pathogen X infection in [Country Y]. It is intended to be conducted before widespread community transmission occurs, that is, within the early phases of the pathogen X epidemic in the country. However, a study can continue for as long as is determined feasible by the country implementing the investigation, particularly if objectives shift to address alternative questions.

#### 2.2.3 Population of interest & study representativeness

The study population is the first few cases of pathogen X and their close contacts in [Country Y]. Index cases will be identified through national line listings/initial case register or other relevant surveillance systems.

However, it is important to note that the early cases are unlikely to be broadly representative of the general population, as identified cases are likely to be from persons with more severe disease that present for healthcare which are more easily detected by a surveillance system. Furthermore, early cases need to satisfy the clinical case definition including the presence of particular exposures (such as international travel, contact with another case) and this may exclude other cases if the pathogen is already circulating in [Country Y].

Where possible, laboratory-confirmed cases in the early epidemic stages should be enrolled (including both symptomatic and asymptomatic cases). If case numbers begin to rise rapidly, the proportion of cases to include could be reduced according to the capacity of [Country Y] (see **Section 2.2.3 CLICK TO VIEW** ). Attempts to follow up all cases in an FFX database can be resource and time intensive.

**Comment:** 

As an example, the United Kingdom's FF100 project for pandemic A(H1N1) influenza ran for 3 months, from April to June 2009, with a total of 392 confirmed cases followed up (20).

In a situation where there are more eligible index cases than can be recruited due to capacity constraints, recruitment could be randomized to reduce the potential for ascertainment biases. It is important that any randomization is considered in relation to geography (such as urban vs rural areas), sex, age, among other factors, to attempt to understand what is occurring at the population-level.

A **screening log** should be used to collect basic information on recruited and non-recruited cases to understand and describe the representativeness of the FFX sample.

# 2.3 Eligibility criteria of cases for the study

#### **Inclusion criteria:**

- The **first few cases of pandemic pathogen X in [Country Y]** should be enrolled according to the case definitions provided in **Box 1**.
- Participating individuals give appropriate informed consent (See Section 2.9 for further details)

# 2.4 Variations to the FFX study

Adapting the FFX study protocol into a longitudinal study may be appropriate to assess questions relating to the development and protectiveness of immunity in a rapidly changing epidemic.

For FFX studies that are extended over longer periods of time, or conducted in later stages of the pandemic, it is advised that they be conducted in geographical locations that are representative of the study country where possible, so that results may be generalizable to the general population.

#### 2.5 Data collection

This FFX protocol calls for the recruitment and follow-up of index cases and their close contacts for a maximum period of 28 days from identification of the index case. **The first day of participation in the FFX investigation will be defined as Day 1.** The duration of follow-up may vary and need to be adapted, depending on the characteristics and transmission dynamics of the pathogen, antibody kinetics and specific public health priorities – see the sampling rationale for more information.

Every effort should be made to include all known **close contacts**, including infants and children, of the index case, to generate the specimen and data sampling time frame for follow-up.

Epidemiological, clinical, virological (if applicable) and serological data will be collected from each participant at multiple times during the study – including surveys at baseline and day 28, symptom diaries from days 2-28 and specimen collection (as per Section 2.5 CLICK TO VIEW and 2.6 CLICK TO VIEW). This information can be obtained through a combination of methods including: face-to-face or telephone interviews/consults with participants (or family members if the case is too ill to be interviewed), self-reporting, interview of health workers and/or review of medical records, self-swabbing and professional/medical specimen collection services.

**Comment:** 

Additional study visits can be included to collect further clinical information and specimens as required. For example, at days 7, 14 and 21.



#### **Toolkit item**

Toolkit items to support this section may include:

- · Data dictionaries
- · Data quality checklist
- · Study team training template

Toolkit components will be available on the WHO website. CLICK TO VIEW

#### 2.5.1 Clinical and epidemiological data collection

Once an index case has been identified and recruited into the investigation, a study visit (if case is hospitalized due to severe illness/isolation purposes or isolating) or phone interview will need to be conducted to identify all close contacts for recruitment into the study. A contact line list can be used to record all close contacts identified (see **Appendix B** CLICK TO VIEW , Contact Line List).

# A summary of the data and specimen collection forms can be found in Table 2 and Figure 3.

For index cases, data will be collected using **Form A1 CLICK TO VIEW** for the baseline visit (day 1), followed by **Form A2 CLICK TO VIEW** for the follow up visit (day 28) to collect relevant sociodemographic and clinical information.

For close contacts, data will be collected using **Form B1 CLICK TO VIEW** for the baseline visit (day 1), followed by **Form B2 CLICK TO VIEW** for the follow up visit (day 28) to collect relevant sociodemographic and clinical information.

Form C CLICK TO VIEW will be used for **all** specimen collection visits (throughout day 1 to day 28) for cases and close contacts.

Form D – symptom diaries (template available in Appendix B of this protocol) will be provided for all cases and close contacts to complete (throughout day 2 to day 28), to record the presence or absence of various signs or symptoms. A proxy may fill out the symptom diaries on behalf of those unable to complete the form themselves. Note that symptoms are recorded on day 1 as part of the baseline form (Form A1 CLICK TO VIEW OR FORM B1 CLICK TO VIEW).

Some aspects to keep in mind are:

contacts found to be infected with pathogen X would be reclassified as confirmed
 cases (dotted line in Figure 3) and follow-up would occur as described in the case
 investigation algorithm. The data collection process may or may not be re-triggered
 in this instance, but would depend on the country resources and type of contact
 (for example, if the contact is a health worker, then a further investigation might be
 warranted to inform public health action).

All **investigation questionnaires can be found in Appendix B CLICK TO VIEW** of this document. Note that the questionnaires may need to be adapted based on the local setting of implementation and the specific characteristics of the pathogen. For example, the required duration of symptom diaries may be shortened to 14 days where pathogen X does not cause prolonged symptoms in cases (i.e., where disease is acute and short in duration, is not known to cause post-viral sequelae after recovery or testing negative) and where chains of infection amongst contacts are not prolonged across the study period.

Figure 3: Case investigation algorithm and summary of data-collection tools

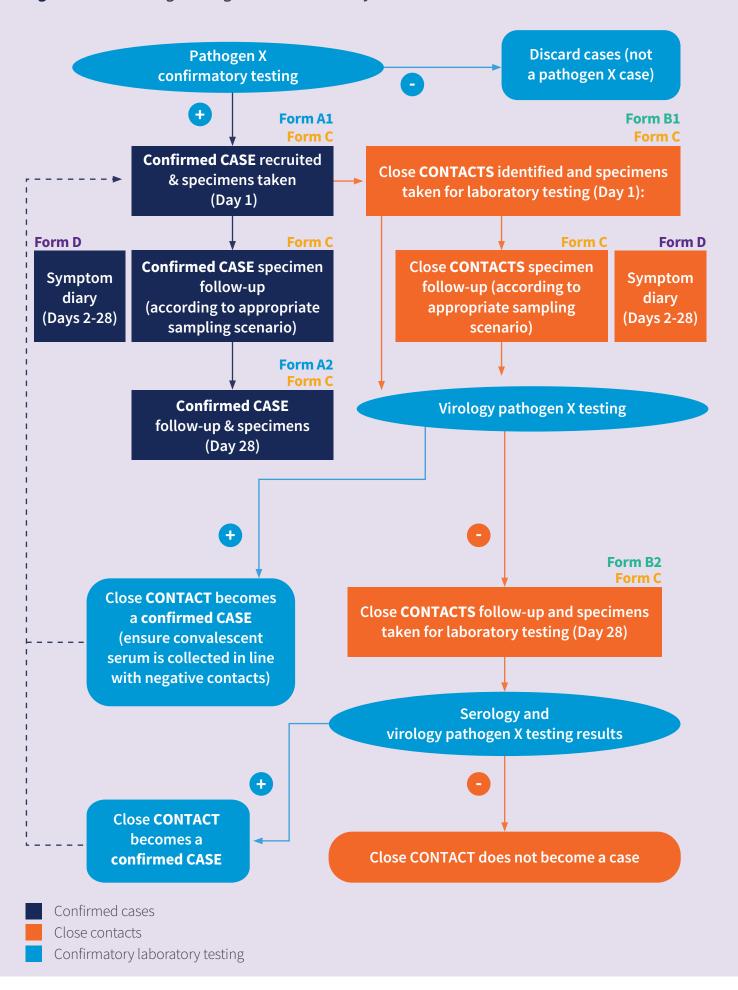


Table 2: Summary of data collection tools

Form number	Purpose of form	Collecting from whom?	When should it be collected?
CASES			
Form A1	Case <b>initial</b> report form	For pathogen X index cases	Day of recruitment into the FFX investigation (as soon as possible after identification of a case) ( <b>Day 1</b> )
Form A2	Case <b>follow-up</b> form	For pathogen X index cases: final outcome	28 days after completion of Form A1 <b>(Day 28)</b> Updates should be obtained regularly, if all the required information is not available at the time of completing this form
CONTACTS			
Form B1	Contact <b>initial</b> reporting form	For contacts of pathogen X index cases	Day of recruitment into the FFX investigation (ideally within 24 hours after identification of the index case) (Day 1)
Form B2	Contact <b>follow-up</b> form	For contacts of pathogen X index cases: final outcome	28 days after completion of Form B1 (Day 28)
CASES & COI	NTACTS		
Form C	Track and summarize all laboratory results (and methods used)	For pathogen X index cases and contacts	This table will need to be filled/updated at <b>each</b> specimen collection time point
Form D: Symptom diary	Record the presence or absence of various signs or symptoms	For pathogen X index cases and contacts	Between <b>days 2 and 28</b> after administration of the initial questionnaire

# 2.6 Specimen collection and transport

The following is intended to guide specimen collection from index cases and their close contacts. Careful planning should be done to ensure arrangements are in place for specimen collection, how to access required PPE and how to arrange safe sample collection and transport of specimens. Sample collection and transport should follow national guidelines.

#### **Comment:**

Guidance on laboratory testing is subject to change, depending on the context of the specific evolution of the epidemic/pandemic, along with the possible development of point of care tests or rapid antigen tests with proven adequate accuracy as they become available.



Toolkit items to support this section may include:

- Standard operating procedure for sample collection
- · Advice for specimen types to be collected

Toolkit components will be available on the WHO website. CLICK TO VIEW

#### 2.6.1 Sampling rationale

This section outlines the rationale for the collection of respiratory and serological specimens from cases and close contacts (Box 4).



#### Box 4. Rationale for specimen collection

In general, the collection of specimens helps to characterise pathogen transmission patterns through:

- Confirming infection in close contacts and classifying participants (Box 3);
- Studying transmission dynamics (infection attack rates, incubation period, infectiousness relative to symptom onset, generation time);
- Determining the extent and fraction of mild or asymptomatic infection;
- Identify symptomatic infections with atypical clinical presentation, and;
- Detecting prolonged viral shedding in cases.

#### Serological specimens will help to:

- Identify seroconversion increase in pathogen specific antibody titres between baseline and convalescent paired samples, in cases and contacts;
- Identify any cases in close contacts not identified by PCR with or without symptoms, and;
- Study transmission dynamics (infection attack rates).

#### Genomic analyses of specimens may help to:

- Conduct advanced analyses of transmission dynamics;
- Strengthen inference on relatedness of infections and geographic spread;
- Quantify genetic diversity, and;
- Identify genetic differences by variants/subtypes/lineages and strengthen inference on phenotypic differences, if any.

The collection of respiratory specimens from (cases and) close contacts should consider:

• The **timing and frequency of sample collection** in relation to the generation interval (see **Figure 1** CLICK TO VIEW ) and the setting of contact to accurately define infection events. Specimen collection should be sufficiently late to allow identification of secondary cases, but early enough to avoid capturing tertiary (or later) cases for estimation of the secondary attack rate;

#### **Comment:**

Characterising pathogens with a short generation time and/or high transmissibility requires respiratory samples to be collected at a higher frequency to identify and observe chains of infection.

The types of respiratory specimen collected (i.e., nasal swabs, nasopharyngeal swab, throat swabs, saliva, sputum, and combinations of these) and the demonstrated sensitivity and specificity of each for detecting pathogen X using RT-PCR or the laboratory method being recommended for pathogen detection (Appendix C CLICK TO VIEW );

#### **Comment:**

In some cases, several specimen types may be indicated.

• How the specimens will be collected (professionally or self-collected). It may be possible for participants to self-collect respiratory specimens. This may reduce staffing burden/costs on study nurses or professional collection services and enable frequent sampling when required per the sampling strategy. For example, self-swabbing has been demonstrated to be a reliable method for influenza and SARS-CoV-2 testing (21), although this is dependent on pre-planning, logistics of transport and the quality of training provided to participants (21, 22).

#### **Comment:**

Investigators may wish to consider self-swabbing to supplement study visits from a research/study nurse or professional specimen collection service. However the first and last study visits should be carried out by study personnel.

The collection of serological specimens from (cases and) close contacts should consider:

• The number of serological specimens. At a minimum, paired samples should be taken at baseline and at day 28 (i.e., at the end of follow-up) to allow time for the maturation and development of the immune response.

#### **Comment:**

Additional serological samples collected during the study period may help to better characterise immune responses in relation to exposure, infection, disease onset, and maturation/development of immunity over time. This may be especially important if pathogen X is novel.

#### **Comment:**

If reliable serology for pathogen X is not available, consider how these specimens will be stored for future testing. Testing can also be performed at a reference laboratory.

#### 2.6.2 Sampling strategy

The sampling strategy for the investigation is presented in this section in Figure 4.

CLICK TO VIEW

Four exemplar scenarios have been selected per Table 3, to account for different plausible generation interval distribution/times of pathogen X as per biological characteristics of known respiratory viruses. These will be henceforth referred to as scenarios 1-4.

**Table 3: Exemplar scenarios of pathogen X** 

Scenario	Pathogen characteristics
1	Mean generation time of 2.6 days (similar to influenza)
2	Mean generation time of 3.6 days (similar to SARS-CoV-2)
3	Mean generation time of 4.6 days
4	Mean generation time of 7 days

Each scenario/strategy involves the mandatory collection of respiratory and serology specimens from **all index cases and all close contacts – irrespective of symptoms** – at determined timepoints.

The early timepoints in each scenario have been selected with the aim of identifying coprimary cases as well as secondary cases as quickly as possible after they are infected, to minimise the risk of misclassification of subsequent tertiary cases as secondary cases. This is especially important when the generation interval is short, as there is a higher likelihood of secondary cases transmitting to tertiary cases prior to recruitment of the index case. Similarly, if testing is not frequent enough or case ascertainment is based on clinical signs and symptoms only, tertiary cases may be misclassified as secondary cases.

#### **Comment:**

Adherence to the harmonised schedules is critical, especially in closed settings where chains of transmission may be rapid.

#### **Comment:**

There is typically a delay between identification of the index case and the recruitment of close contacts into the investigation. The scenarios prioritise early sample collection to account for the potential for transmission to already have occurred prior to recruitment.

A final respiratory and serological specimen is mandatory at the end of follow-up to study the immune response and inform estimates of overall attack rates and any prolonged viral shedding.

In addition to this generic protocol, the chosen sampling strategy in the FFX investigation in [Country Y] should be informed by current knowledge of the biological and epidemiological characteristics of pathogen X (e.g., the incubation period, latent period) and the expected time delays associated with testing, notification, and recruitment of a case). The strategy should be readily adapted/flexible to new information as more becomes known about pathogen X over time.

Where appropriate, **other specimens** (e.g., oral fluid, urine, faeces) may be collected, according to clinical presentation and observed patterns of viral shedding upon infection.

#### **Comment:**

The onset of acute respiratory illness in contacts may prompt further testing, however this must be **in addition** to the mandatory components of the proposed sampling schedule and not a replacement for standardised testing.

#### **Comment:**

Countries undertaking FFX investigations may wish to modify their sampling requirements depending on their specific or additional study objectives and availability of resources. We strongly advise that coordinators consult the sampling rationale and statistical analysis plan prior to adapting or modifying this strategy.

Figure 4: Sampling strategy for scenarios 1-4

,	tment	1	2	3 4	5	6 7	8	9	10 11	12ء	13	14	15	L6 1	7 18	19	20	21	22	23	24	25	26 2	7 2
Study visit																								
Respiratory sampl	es							Ti																
Blood/Serum sam	ple																							
Other specimens (if	f/where relevant)																							
Scenario 2: Me	ean generatio	n t	time	e of	3.6	6 da	ys (	sin	nilar	to S	SAR!	s-c	oV-	2)										
Day since recrui	tment	1	2	3 4	5	6 7	8	9	10 11	12	13	14	15	L6 1	7 18	19	20	21	22	23	24	25	26 2	7 2
Study visit																								
Respiratory sampl	es																							
Blood/Serum sam	ple																							
Other specimens (if	f/where relevant)									,														
Scenario 3: Me	ean generatio	n t	time	e of	4.6	6 da	ys																	
Day since recrui	tment	1	2	3 4	5	6 7	8	9	10 11	12	13	14	15	16 1	7 18	19	20	21	22	23	24	25	26 2	7 :
Study visit																								
Respiratory sampl	es																							
Blood/Serum sam	ple																							
Other specimens (if	f/where relevant)																							
Scenario 4: Me	ean generatio	n t	time	e of	7 0	days																		
Day since recrui	tment	1	2	3 4	5	6 7	8	9	10 11	12	13	14	15	L6 1	7 18	19	20	21	22	23	24	25	26 2	7 2
Study visit																								
Respiratory sampl	es																							
Blood/Serum sam	ple																							
	f/where relevant)																							

#### 2.6.3 Specimen collection and transport

Appropriate PPE should be worn when specimens are being collected from index cases and all contacts from professional/health-care specimen collection services (23). All those involved in collecting and transporting specimens should be trained in the safe handling of infectious substances and infectious spill decontamination procedures. For details regarding the collection and transport of samples, please refer to the case management algorithm and laboratory guidance in the country, and to WHO laboratory guidance (24).

For each biological sample collected, the time of collection, the conditions for transportation and the time of arrival at the laboratory should be recorded. Specimens should ideally be collected within 3 days of the onset of clinical symptoms and reach the laboratory as soon as possible after collection. If the specimen is not likely to reach the laboratory and to be processed within 48 hours, it should be frozen, preferably at –80 °C, and shipped on dry ice as per applicable WHO guidance (24). It is, however, important to aliquot each sample to avoid the deleterious effects of such handling on the infectivity of virus and the yield of genetic material. The storage of respiratory and serum specimens in domestic frost-free freezers should be avoided, owing to their wide temperature fluctuations. Serum should be separated from whole blood and can be stored at 4 °C for one week and shipped at 4 °C or frozen to –20 °C or lower and shipped on dry ice.

Transport of specimens within national borders should comply with applicable national regulations. International transport of specimens should follow applicable international regulations as described in the WHO Guidance on regulations for the transport of infectious substances 2021–2022 (25).

# 2.7 Laboratory evaluations

# 2.7.1 Laboratory analysis

#### **Comment:**

Guidance on laboratory testing is preliminary and may change, depending on the characteristics of pathogen X, the availability of laboratory tests for pathogen X and the context of the specific evolution of the epidemic/pandemic.

Laboratory testing guidance of [Country Y] for pathogen X can be found in **Appendix xx** (country to insert)



# **Toolkit item**

Toolkit items to support this section may include:

• Protocols/standard operating procedures

Toolkit components will be available on the WHO website. CLICK TO VIEW

# 2.8 Data management generalities

The FFX data system will be maintained centrally by [cite institution/body/person(s) in the country].



# **Toolkit item**

Toolkit items to support this section may include:

- Data collection/entry guidance
- Data transfer
- · Data quality checklist
- Data dictionaries
- Data cleaning
- Data security

Toolkit components will be available on the WHO website. CLICK TO VIEW

# 2.9 Ethical considerations

National and local ethical requirements must be followed. Ethical approval should be sought as per individual country requirements as ethical requirements will vary by country. In some countries, this investigation may fall under public health surveillance (emergency response) acts and may not require ethical approval from an institutional review board. It will be upon national authorities to advise on the specific requirements for the investigation in [Country Y].

It is important that, wherever possible, ethics pre-approval is sought in advance of a pandemic to reduce time to activation of Respiratory Investigations and Studies in accordance with local, regional and national authorities. This will ensure that investigations are implemented when necessary and with minimal delay.

For further information on the ethical considerations of importance to public health surveillance can be found in key WHO guidance:

- Guidance for managing ethical issues in infectious disease outbreaks (who.int) (26) CLICK TO VIEW
- WHO guidelines on ethical issues in public health surveillance (27) CLICK TO VIEW



Toolkit items to support this section may include:

- Ethical exemption/clearance letter templates
- Links to key WHO guidance on ethical considerations of importance to public health surveillance
- Template consent and assent forms

Toolkit components will be available on the WHO website. CLICK TO VIEW

#### 2.9.1 Informed consent and assent

The purpose of the investigation will be explained to all individuals willing to participate, before the start of the investigation. For all investigation activities not included in routine public health management, informed consent and assent may be required. This will depend on the country's national ethical requirements. Informed consent will seek relevant approvals for the collection of all data and specimens for the purposes of this investigation as determined by [Country Y].

#### • Consent for:

Adults; and

Children under the legal age of consent (usually 18 years, but will vary from country to country) from a parent or legal guardian.

#### • **Assent** from:

Children and adolescents under the legal age of consent, but who can understand the implications of informed consent and go through the necessary procedures. This is usually children over the age of 12 to 13 years, but this will vary by country. A consent form from a parent or legal guardian will be collected in addition.

#### **Comment:**

The age of consent may vary by country. Check the requirements of local, regional or national authorities.

# **Comment:**

If older adults are being included, assessment of cognitive function (affecting ability to consent) could be included, and consent of legal guardians might be required for adults with cognitive decline if these individuals are to be considered for inclusion.

**Template informed consent and assent forms** will be included in the supporting FFX investigation toolkit.

All eligible individuals, regardless of whether or not they are well or unwell should be able to participate in the investigation. For individuals who lack the decisional capacity to consent at the time of the investigation, consent/assent by proxy (parent/ legal guardian/ spouse/ family member) may be considered so as to not unduly exclude individuals from participating in the investigation.

The processes related to withdrawal of a participant need to be described both in the protocol and in the information given to the participant at the time of enrolment. In this description it must be made clear that a participant can withdraw from the investigation, without justification, at any time by informing one of the members of the investigation team. The contact details of one of the members of the investigation need to be provided in the information for the participant. If any participant decides to withdraw from the investigation, the samples collected and data should be discarded, except if the participant indicates that these can be kept for the purpose of conducting the investigation, or for future studies of other infectious pathogens (See Section 2.9.5 CLICK TO VIEW ).

Informed consent will seek approval to collect blood, respiratory samples, any other relevant samples and epidemiological data for the intended purpose of this investigation. It may also seek approval that samples may be shipped outside of the country for additional testing; and that samples may be stored and used for future public health needs, in accordance with national laws and regulations. Additional detail in the consent/ assent forms may be needed, according to national laws and regulations, if the investigation calls for storage and future use of samples. See further details in Section 2.9.5.

#### **Comment:**

Participants or their parent/legal guardian/spouse will be informed about the test results, with an explanation of the interpretation and implications of the test results.

# 2.9.2 Risks and benefits for subjects

This investigation poses minimal risk to participants, involving the collection of a small amount of blood and respiratory specimens. The direct benefit to the participant is the possibility for early detection of pandemic pathogen X infection, which would allow for appropriate monitoring and treatment for themselves and their close contacts. The primary benefit of the investigation is indirect, in that data collected will help improve and guide efforts to understand transmission of pandemic pathogen X and inform appropriate public health responses.

In terms of treatment of cases, case management will be facilitated by early detection of the disease and will follow national guidance, but the investigators may or may not be directly involved in clinical management of patients. Processes on how cases will be referred for medical care, as well as details on provisions of care as part of the investigation will need to be detailed.

There will be no incentives while participating in this investigation, but participants will be provided relevant and updated health advice to reduce the risk of transmission and severe outcomes where possible.

# 2.9.3 Reporting of serious adverse events, including death of a participant

Any serious adverse event<sup>5</sup>, including death, of a participant during the investigation period, needs to be immediately (within 24h) reported to the Principal Investigator and the institution responsible for the investigation. The contact details for reporting serious adverse events needs to be provided to each member of the investigation team.

In accordance with national regulations, any serious adverse event, may also have to be reported to the local ethical review committee, if the adapted protocol was not deemed exempt from local ethical review committee.

# 2.9.4 Confidentiality

Participant confidentiality will be maintained throughout the investigation. All subjects who participate in the investigation will be assigned a unique identification number by the investigation team, for the labelling of questionnaires and clinical specimens. The link of this identification number to individuals will be maintained by the investigation team and will not be disclosed elsewhere.

Data and specimens must be securely stored. If the data are shared by the implementing organization with WHO or any agency or institution providing support for data analysis, the shared data will include only the investigation identification number and not any personally identifiable information. Data sharing outside [Country Y] must be managed according to national laws and regulations, as appropriate.

# **Comment:**

The investigators will need to describe how data and specimens will be securely stored, the duration of storage and the destruction of data and specimens at the end of the duration of storage, in accordance with national laws and regulations.

<sup>5</sup> An adverse event can be defined as: An injury related to medical management, in contrast to complications of disease(34).

Article 45 of the International Health Regulations (2005) (IHR) describes the "treatment of personal data" (28). Personally identifiable data collected under the IHR should be kept confidential and processed anonymously, as required by national law. However, such data may be disclosed for assessments and management of public health risks, provided the data are processed fairly and lawfully.

# 2.9.5 Future use of samples

The investigators may decide on potential future use of specimens and the time-frame for their destruction and seek appropriate approvals for this. If this is the case, the investigators will need to provide more specific information on potential future use of specimens and the time-frame for their destruction, including in the information for the participant and the informed consent/assent form. Additional consent forms may need to be developed by the country, to comply with national laws and regulations.

# 2.9.6 Prevention of pathogen X infection among investigation personnel

All personnel involved in the investigation visits must be trained in procedures for infection prevention and control (as determined by national or local guidelines) (29) when collecting data or specimens in proximity to cases and close contacts. These procedures should include proper hand hygiene and the correct use of surgical or respiratory face masks, to minimize their risk of infection.

WHO technical guidance on infection prevention and control specific to pathogen X will be published on the WHO website.

# 3. Statistical analyses



# 3.1 Sample size

The sample size of the investigation will be determined by the number of contacts within each sphere of activity of the index cases and assumptions made relating to the transmissibility of pathogen X. **Every effort should be made to include all contacts of the index case, to improve the precision and reduce bias in parameter estimates.** However as noted in Section 1.2, these investigations are resource intensive so it may be best to focus initially on the follow-up of **household and health-worker contacts**.

For further details please refer to the Toolkit item, Statistical Analysis Plan, available on the WHO website.

Sample sizes can be calculated using **statistical formulas or tools** available online (e.g., http://www.openepi.com/Menu/OE\_Menu.htm CLICK TO VIEW ) or in standard statistical packages.

# 3.2 Plan of analysis

FFX investigations will not be able to answer every question we have about pathogen X infection, but they will contribute key data in the early stages of an epidemic, which can inform an appropriate public health response. Other protocols for investigations adapted for pathogen X can assist in providing supplementary data to help with the calculation of key epidemiological parameters.

Prior to commencing the investigation, an analysis plan should be developed incorporating the study objectives, definitions and planned analysis to address each objective. Pre-specifying the analysis plan will help to ensure that all relevant data are being collected. Please refer to the WHO Statistical Analysis Plan as part of the supporting **Toolkit**.

A descriptive analysis (time, place, person) of the FFX should provide preliminary insight into the demographics of individuals infected with pathogen X, as well as, severity, the clinical spectrum and course of disease – for example, the initial population groups most affected, by age and underlying risk factors. See Figure 5 and Section 4 CLICK TO VIEW for more details.

**A statistical analysis** (epidemiological parameters estimation) of the FFX data will provide estimates of the transmissibility and severity of pathogen X – for example, the secondary attack rate or hospitalization/fatality rate by setting, age and other risk factors. See **Figure 5** and **Section 4 CLICK TO VIEW** for more details.

Additional complex analyses can be conducted using the FFX forms/questionnaires and specimens generated as described in Figure 5. These require more computationally intensive analysis methods, i.e., mathematical modelling approaches.

**Genomic analysis** of the specimens generated through this investigation can help provide a detailed insight into the origin of the pandemic; monitor the potential spread of antiviral resistance; and identify transmission chains using the confirmed case as a potential origin (by comparing the relatedness of two virus isolates), which in turn will help with estimation of the basic reproduction number. Genomic analyses can be useful for determination of the extent of community transmission that is occurring in the early stages of the pandemic and whether the strain was locally acquired or imported from another region.



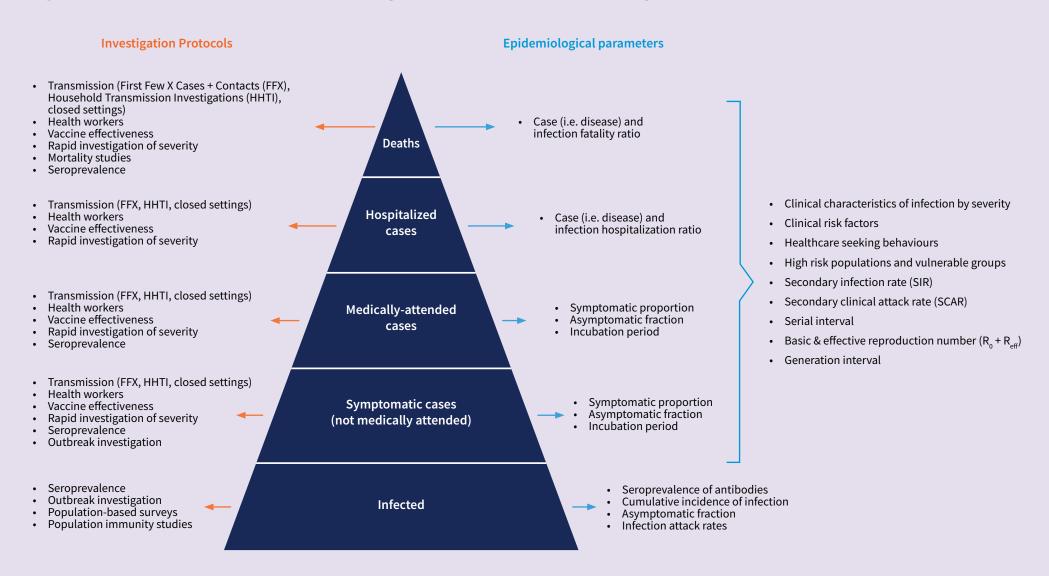
# **Toolkit item**

Toolkit items to support this section may include:

- Template statistical analysis plans
- · Data analysis scripts
- · SOP for interpretation
- · Reporting guidelines for outcomes

Toolkit components will be available on the WHO website. CLICK TO VIEW

Figure 5: Disease pyramid with associated epidemiological parameters to be estimated through implementation of Unity Studies protocols



Data sharing and reporting of findings

# 4.1 Data sharing

Data from these WHO Respiratory Investigations and Studies may be **pooled and aggregated across multiple sites** by WHO if the data are collected in a consistent manner, to increase analytic power and improve precision in estimates of severity and transmissibility. The pooling of this data will depend on how transmission dynamics vary between countries. If the data are shared by the implementing organization or country, with WHO or with any agency or institution providing support for data analysis, data shared will include only the investigation identification number and not any personally identifiable information.

Any data sharing must also be covered by any ethical approvals, where relevant, or national regulations.



# **Toolkit item**

Toolkit items to support this section may include:

- · Data sharing agreement templates
- · Platforms for data sharing
- Standardized formats/templates for data sharing

Toolkit components will be available on the WHO website. CLICK TO VIEW

# 4.2 Reporting of findings

Any investigation of this nature should include reporting on the following information, stratified by relevant demographic, time and place characteristics:

- the number of cases (symptomatic and asymptomatic) and number of close contacts included:
- the number of pathogen X cases among the close contacts;
- the number of symptomatic and asymptomatic close contacts;
- the number of close contacts with serological evidence of pathogen X infection:
- · screening log of recruited and non-recruited FFX cases, and;
- Parameter estimates with uncertainty.

In addition, further details regarding the study should be reported, including:

- the timing of the study;
- inclusion / exclusion criteria of cases and contacts and any loss to follow up;
- case and close definitions used:
- context of the study, e.g., community incidence/transmission, geographical spread, any pharmaceutical/non-pharmaceutical interventions where applicable;
- statistical methods used to calculate estimates.

**Timely dissemination** of the results of this investigation is critical to update guidance and inform local, national and international public health responses and policies for infection prevention and control. WHO needs to be able to adapt WHO global recommendations in a timely manner. Countries are encouraged to share with WHO in line with IHR requirements and in a confidential manner, any early findings, especially if they will impact WHO global guidance.

It is also important to fully **document the investigation design**, including but not limited to the definition of close contacts; the approach to ascertainment of primary cases and secondary cases (including any inclusion and exclusion criteria); the duration of follow-up; and the laboratory methods used to ensure that data can be pooled to increase the precision of epidemiological parameter estimates.

Information should be collected in a **standardized format** according to the questionnaires and tools in this generic protocol, to assist with data harmonization and comparison of results (see forms in **Appendix B** CLICK TO VIEW ).



# **Toolkit item**

Toolkit items to support this section may include:

- Standard criteria for reporting results
- Templates for presentations and publications
- · Training on messaging to different audiences
- Technical briefs

Toolkit components will be available on the WHO website. CLICK TO VIEW

# 4.3 Science translation for decision makers

Clear and timely reporting and communication of results is necessary to support policy/ decision makers.



# **Toolkit item**

Toolkit items to support this section may include:

- Training on messaging to different audiences
- Policy makers questions that will be answered by study objectives
- Science translation materials

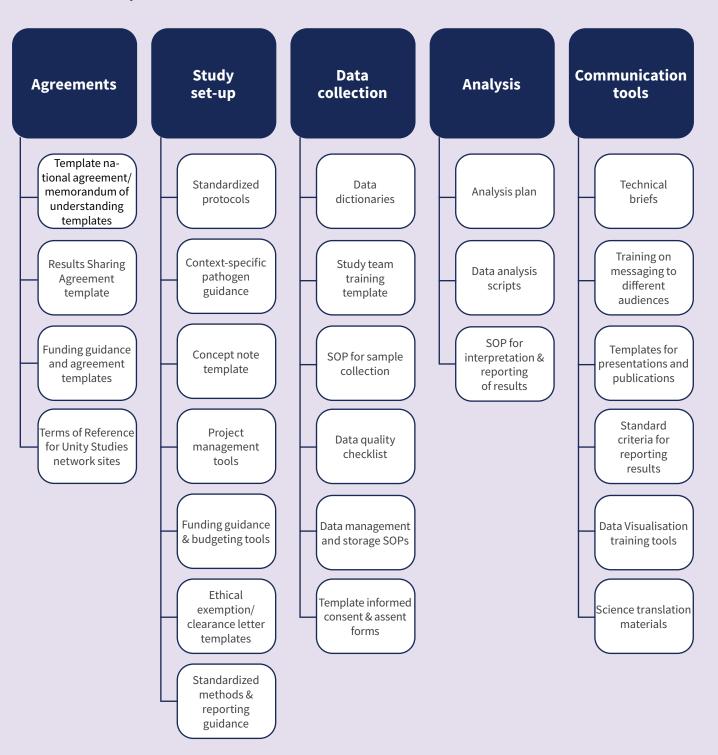
Toolkit components will be available on the WHO website. CLICK TO VIEW

# 5. Protoco Toolkit **Protocol**

# 5.1 Protocol Toolkit

The timely and quality implementation of this protocol should be supported by the use of toolkits developed by WHO and implementing partners. These will include components such as pre-planned agreements, study set-up resources, data collection, analysis and communication tools as shown in **Figure 6** and highlighted throughout the protocol.

**Figure 6:** Proposed toolkit components to support quality implementation of Investigations and Studies protocols



References 6.

- 1. Consortium for the Standardization of Influenza Seroepidemiology (CONSISE). CONSISE Home [Internet]. 2023 [cited 2023 Jul 12]. Available from: https://consise.tghn.org/
- World Health Organization. Coronavirus disease (COVID-19) technical guidance: The Unity Studies: Early Investigation Protocols [Internet]. 2023 [cited 2023 Jul 12]. Available from: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/early-investigations
- 3. World Health Organization. Middle East Respiratory Syndrome Outbreak Toolbox [Internet]. 2022 [cited 2023 Jul 12]. Available from: https://www.who.int/emergencies/outbreak-toolkit/disease-outbreak-toolboxes/mers-outbreak-toolbox
- 4. World Health Organization. Final report of the IHR Review Committee (as presented to 64th WHA) [Internet]. 2011 [cited 2023 Jul 12]. Available from: https://www.who.int/teams/ihr/ihr-review-committees/h1n1
- 5. World Health Organization. Global Influenza Programme [Internet]. 2023 [cited 2023 Jul 12]. Available from: https://www.who.int/teams/global-influenza-programme
- 6. World Health Organization. Pandemic Influenza Preparedness (PIP) Framework [Internet]. 2023 [cited 2023 Jul 12]. Available from: https://www.who.int/initiatives/pandemic-influenza-preparedness-framework
- 7. World Health Organization. WHO Zika virus research agenda [Internet]. 2016 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/WHO-ZIKV-PHR-16.1
- 8. Bergeri I, Whelan MG, Ware H, Subissi L, Nardone A, Lewis HC, et al. Global SARS-CoV-2 seroprevalence from January 2020 to April 2022: A systematic review and meta-analysis of standardized population-based studies. PLoS Med [Internet]. 2022 Nov 1 [cited 2023 Jul 12];19(11). Available from: https://pubmed.ncbi.nlm.nih.gov/36355774/
- 9. Lewis HC, Ware H, Whelan M, Subissi L, Li Z, Ma X, et al. SARS-CoV-2 infection in Africa: a systematic review and meta-analysis of standardised seroprevalence studies, from January 2020 to December 2021. BMJ Glob Health [Internet]. 2022 Aug 23 [cited 2023 Jul 12];7(8). Available from: https://pubmed.ncbi.nlm.nih.gov/35998978/
- Hennessey K, Pezzoli L, Mantel C. A framework for seroepidemiologic investigations in future pandemics: insights from an evaluation of WHO's Unity Studies initiative. Health Res Policy Syst [Internet]. 2023 Dec 1 [cited 2023 Jul 12];21(1). Available from: https:// pubmed.ncbi.nlm.nih.gov/37194007/
- 11. World Health Organization. Global Influenza Strategy 2019–2030 [Internet]. 2019 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/9789241515320

- 12. World Health Organization. Pandemic influenza preparedness framework: partnership contribution high-level implementation plan III 2024-2030 [Internet]. 2023 [cited 2023 Jul 14]. Available from: https://www.who.int/publications/i/item/9789240070141
- 13. World Health Organization. Mosaic Respiratory Surveillance Framework [Internet]. 2023 [cited 2023 Jul 13]. Available from: https://www.who.int/initiatives/mosaic-respiratory-surveillance-framework/
- 14. Cauchemez S, Bosetti P, Cowling BJ. Managing sources of error during pandemics. Science [Internet]. 2023 Feb 3 [cited 2023 Jul 13];379(6631):437–9. Available from: https://pubmed.ncbi.nlm.nih.gov/36730404/
- 15. World Health Organization. Non-pharmaceutical public health measures for mitigating the risk and impact of epidemic and pandemic influenza [Internet]. 2019 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/non-pharmaceutical-public-health-measuresfor-mitigating-the-risk-and-impact-of-epidemic-and-pandemic-influenza
- 16. Pan American Health Organization. Norms and Standards in Epidemiology: Case Definitions. Epidemiol Bull. 1999;20(2):13.
- 17. Price DJ, Spirkoska V, Marcato AJ, Meagher N, Fielding JE, Karahalios A, et al. Household transmission investigation: Design, reporting and critical appraisal. Influenza Other Respir Viruses. 2023;17(6).
- 18. Lau LLH, Nishiura H, Kelly H, Ip DKM, Leung GM, Cowling BJ. Household transmission of 2009 pandemic influenza A (H1N1): a systematic review and meta-analysis. Epidemiology [Internet]. 2012 Jul [cited 2023 Jul 12];23(4):531–42. Available from: https://pubmed.ncbi.nlm.nih.gov/22561117/
- 19. Giesecke J. Primary and index cases. 2014 [cited 2023 Jul 12]; Available from: http://www.
- 20. McLean E, Pebody RG, Campbell C, Chamberland M, Hawkins C, Nguyen-Van-Tam JS, et al. Pandemic (H1N1) 2009 influenza in the UK: clinical and epidemiological findings from the first few hundred (FF100) cases. Epidemiol Infect [Internet]. 2010 Nov [cited 2023 Jul 12];138(11):1531–41. Available from: https://pubmed.ncbi.nlm.nih.gov/20594381/
- 21. Ip DKM, Schutten M, Fang VJ, Fung ROP, Dutkowski RT, Chan KH, et al. Validation of self-swab for virologic confirmation of influenza virus infections in a community setting. J Infect Dis [Internet]. 2012 Feb 15 [cited 2023 Jul 12];205(4):631–4. Available from: https://pubmed.ncbi.nlm.nih.gov/22198963/

- 22. Elliot AJ, Powers C, Thornton A, Obi C, Hill C, Simms I, et al. Monitoring the emergence of community transmission of influenza A/H1N1 2009 in England: a cross sectional opportunistic survey of self sampled telephone callers to NHS Direct. BMJ [Internet]. 2009 Sep 12 [cited 2023 Jul 12];339(7721):619. Available from: https://pubmed.ncbi.nlm.nih.gov/19713236/
- 23. World Health Organization. Clinical care of severe acute respiratory infections Tool kit [Internet]. 2022 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/clinical-care-of-severe-acute-respiratory-infections-tool-kit
- 24. World Health Organization. Laboratory biosafety manual, 4th edition [Internet]. 2020 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/9789240011311
- 25. World Health Organization. Guidance on regulations for the transport of infectious substances 2021-2022 [Internet]. 2021 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/9789240019720
- 26. World Health Organization. Guidance for managing ethical issues in infectious disease outbreaks [Internet]. 2016 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/9789241549837
- 27. World Health Organization. WHO guidelines on ethical issues in public health surveillance [Internet]. 2017 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/who-guidelines-on-ethical-issues-in-public-health-surveillance
- 28. World Health Organization. International Health Regulations (2005) Third Edition [Internet]. 2016 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/9789241580496
- 29. World Health Organization. Infection prevention and control of epidemic- and pandemic-prone acute respiratory infections in health care [Internet]. 2014 [cited 2023 Jul 13]. Available from: https://www.who.int/publications/i/item/infection-prevention-and-control-of-epidemic-and-pandemic-prone-acute-respiratory-infections-in-health-care
- 30. World Health Organization. Investigation of cases of human infection with Middle East respiratory syndrome coronavirus (MERS-CoV): interim guidance [Internet]. 2015 [cited 2023 Jul 14]. Available from: http://www.who.int/iris/handle/10665/178252
- 31. World Health Organization. Interim WHO surveillance recommendations for human infection with avian influenza A(H7N9) virus [Internet]. 2013 [cited 2023 Jul 14]. Available from: http://www.who.int/influenza/human\_animal\_interface/influenza\_h7n9/InterimSurveillanceRecH7N9\_10May13.pdf?ua=1

- 32. World Health Organization. Collecting, preserving and shipping specimens for the diagnosis of avian influenza A(H5N1) virus infection: guide for field operations [Internet]. 2006 [cited 2023 Jul 14]. Available from: http://www.who.int/csr/resources/publications/surveillance/WHO\_CDS\_ EPR\_ARO\_2006\_1/en/index.html
- 33. World Health Organization. Protocol to investigate non-seasonal influenza and other emerging acute respiratory diseases [Internet]. 2018 [cited 2023 Jul 14]. Available from: https://apps.who.int/iris/handle/10665/275657
- 34. World Health Organization. WHO Draft Guidelines for Adverse Event Reporting and Learning Systems [Internet]. 2005 [cited 2023 Jul 19]. Available from: https://www.who.int/publications/i/item/WHO-EIP-SPO-QPS-05.3

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**Appendix** 

# Appendix A: Comparison between the features and complementarity of the main respiratory pathogens of pandemic potential investigation protocols

	The First Few X cases and contacts (FFX) investigation protocol for respiratory pathogens of pandemic potential	Household transmission investigation protocol for respiratory pathogens of pandemic potential	Closed setting transmission investigation protocol for respiratory pathogens of pandemic potential
Population	The First Few X cases of pathogen X and their close contacts in the general population	Household close contacts of index cases of pathogen X (smaller epidemiological unit than FFX)	Contacts of index cases in the closed setting (larger epidemiological unit)
Aim	Transmission dynamics, severity and clinical spectrum, in a proxy of the general population	Transmission dynamics, severity and clinical spectrum, in household settings	Transmissibility- chains of infection in closed settings such as schools, hospitals and army barracks
Potential output and analysis	Transmission dynamics, severity and clinical spectrum, through estimates of, primarily:  • the secondary infection rate (SIR) and secondary clinical attack rate of pathogen X, and by key factors such as setting age and sex  • the clinical presentation of pathogen X infection and course of associated disease  • the symptomatic & asymptomatic proportions of pathogen X cases  • preliminary case and infection hospitalization and fatality ratios.  and secondarily:  • the serial interval of pathogen X  • duration of viral shedding (if samples are taken at higher frequency and adequate resources are available)  • identification of possible routes of transmission including possible animal/human transmission  • risk and/or protective factors for transmission or severe disease  Advance related objectives:  • the basic reproduction number (R <sub>of</sub> ) of pathogen X  • the effective reproductive number (R <sub>eff</sub> ) of pathogen X	<ul> <li>Key data to complement and reinforce the findings of FFX, through estimates of, primarily:</li> <li>the secondary infection rate (SIR) and secondary clinical attack rate (SCAR) of pathogen X in households, and by key factors such as age and sex</li> <li>the clinical presentation of pathogen X infection and course of associated disease</li> <li>the symptomatic &amp; asymptomatic proportions of pathogen X cases</li> <li>preliminary case and infection hospitalization ratios</li> <li>and secondarily:</li> <li>the serial interval of pathogen X</li> <li>duration of viral shedding (if samples are taken at higher frequency and adequate resources are available)</li> <li>risk and/or protective factors for transmission or severe disease</li> <li>Advance related objectives:</li> <li>the incubation period of pathogen X</li> </ul>	<ul> <li>Key data to complement and reinforce the findings of FFX through estimates of, primarily:</li> <li>the overall infection and clinical attack rate of pathogen X, and by key factors such as age and sex</li> <li>the secondary infection rate (SIR) and secondary clinical attack rate of pathogen X infection overall, and by key factors such as setting age and sex</li> <li>the clinical presentation of pathogen X infection and course of associated disease</li> <li>the symptomatic &amp; asymptomatic proportions of pathoger X cases</li> <li>preliminary case and infection hospitalization and fatality ratios.</li> <li>and secondarily:</li> <li>the serial interval of pathogen X</li> <li>risk and/or protective factors for transmission or severe disease</li> <li>Advance related objectives:</li> <li>the basic reproduction number (R<sub>of</sub>) of pathogen X</li> <li>the effective reproductive number (R<sub>eff</sub>) of pathogen X</li> <li>the incubation period of pathogen X</li> <li>the generation interval of pathogen X</li> </ul>

	The First Few X cases and contacts (FFX) investigation protocol for respiratory pathogens of pandemic potential	Household transmission investigation protocol for respiratory pathogens of pandemic potential	Closed setting transmission investigation protocol for respiratory pathogens of pandemic potential
Duration	Recruitment and follow-up of index cases and their close contacts for a maximum period of 28 days from identification of the index case.	Recruitment and follow-up of index cases and their household contacts for a maximum period of 28 days from identification of the index case.	Recruitment and follow-up of index cases and their close contact in a closed setting for a maximum period of 28 days from laboratory confirmation of the index case.
		Study enrolment could be extended as far as desired; however, the most valuable period in order to use data for targeted public health action is in the early phases of the epidemic (first 2–3 months).	Enrolment could be extended as far as desired, however the most valuable period in order to use data for targeted public health action is in the early phases of the epidemic/ pandemic (first 2-3 months).
Start of the investigation	To be initiated in the first days after the identification of a case of pathogen X in [Country Y].	Prospective study, ideally before widespread community transmission occurs, within 2-3 months after identification of initial cases.	Prospective study, ideally before widespread community transmission occurs, within 2-3 months after identification of initial cases.
Recruitment	The first few cases of pathogen X in [Country Y], and their close contacts, will be first few participants to be recruited.	Household contacts of index cases of pathogen X.	Contacts within closed settings will be enrolled.
Minimum data and specimens to be obtained from participants	Epidemiological, clinical, virological (if applicable) and serological and Day 28, symptom diaries from Day 2-28 and specimen collection		tiple times during the study – including surveys at baseline

# **Appendix B: Questionnaires and guidance**

The First Few X cases and contacts (FFX) investigation protocol for respiratory pathogens of pandemic potential

#### **FOR CASES**

- Contact Line List
- Form A1: Case initial report form for pathogen X index cases (Day 1)
- Form A2: Case follow-up form for pathogen X index cases (Day 28)

#### **FOR CONTACTS**

- Form B1: Contact initial reporting form for close contacts of pathogen X cases (Day 1)
- Form B2: Contact follow-up reporting form for close contacts of pathogen X cases (Day 28)

# **FOR CASES AND CONTACTS**

- Form C: Specimen collection and laboratory results
- Form D: Symptom diary for all participants pathogen X index cases and close contacts

# **Contact Line List**

Please copy and transpose the following fields into an Excel spreadsheet and use to populate the Contact Line List. Alternatively use appropriate electronic methods to capture this information. To be completed at the initial visit (day 1) with an index case.

Index Case ID/Cluster number
Contact ID Number (C)
First Name
Family Name
Sex (M/F/Unknown)
DOB (dd/mm/yyyy)
Telephone number
Type of contact (Household/health worker/other)

The First Few X cases and contacts (FFX) investigation template protocol for respiratory pathogens with pandemic potential

# 1. For cases

Form A1: Case initial reporting form – for pathogen X index cases (Day 1)			
Unique Case ID/Cluster number (if applicable):			
1. Current status			
☐ Alive ☐ Dead ☐ Unknown/lost to follow-up			
2. Data collector information			
Name of data collector			
Data collector institution			
Data collector telephone number			
Data collector email			
Form completion date (dd/mm/yyyy)	//		
·			
3. Interview respondent information (if the person	providing the information is not the index case)		
First name			
Family name			
Sex	☐ Male ☐ Female ☐ Non-binary ☐ Not known		
Date of birth (dd/mm/yyyy)	// Unknown		
Relationship to case			

Respondent address

Telephone (mobile) number

4. Patient identifier information	
First name*	
Family name*	
Sex	☐ Male ☐ Female ☐ Non-binary ☐ Not known
Date of birth (dd/mm/yyyy)*	// Unknown
Telephone (mobile) number	
Age (years, months)	years months  Unknown
Email	
Address*	
National social number/identifier (if applicable)*	
Country of residence	
Nationality	
Case occupation (specify location/facility)	☐ Health care worker ☐ Working with animals ☐ Health laboratory worker ☐ Other, specify:  For each occupation, please specify location or facility:
Ethnicity (optional)	☐ Arab ☐ Black ☐ East Asian ☐ South Asian ☐ West Asian ☐ Latin American ☐ White ☐ Aboriginal/First Nations ☐ Other: ☐ Unknown
Is this case part of an institutional outbreak (e.g., aged care facility, hospital, group home)?	☐ Yes ☐ No ☐ Unknown If Yes, specify:
Name of institution involved in outbreak if appropriate	
and should be updated by Country Y according to national guidelines.	ormation and must be kept confidential, however these may vary by country
5a. Patient symptoms (from onset of symptoms)	
Date of first symptom onset (dd/mm/yyyy)	// ☐ No symptoms ☐ Unknown
Fever (≥38 °C) or history of fever since disease onset	☐ Yes ☐ No ☐ Unknown  If Yes date (dd/mm/yyyy): / /

5b. Respiratory symptoms		
Sore throat	☐Yes ☐No ☐Unknown	
	If Yes, date (dd/mm/yyyy):/	
Runny nose	☐ Yes ☐ No ☐ Unknown	
Carrel	If Yes, date (dd/mm/yyyy):/	
Cough	☐ Yes ☐ No ☐ Unknown  If Yes to cough, productive, ☐ Yes ☐ No ☐ Unknown	
	If Yes to cough, dry, ☐ Yes ☐ No ☐ Unknown	
Charles	If Yes, date (dd/mm/yyyy):/	
Shortness of breath	☐ Yes ☐ No ☐ Unknown If Yes, date (dd/mm/yyyy)://	
5c. General symptoms	133331	
Chills	☐ Yes ☐ No ☐ Unknown	
Anosmia (loss of smell)	☐ Yes ☐ No ☐ Unknown	
Ageusia (loss of taste)	☐Yes ☐No ☐Unknown	
Vomiting	☐ Yes ☐ No ☐ Unknown	
Nausea	☐Yes ☐No ☐Unknown	
Diarrhoea	☐Yes ☐No ☐Unknown	
Headache	☐ Yes ☐ No ☐ Unknown	
Rash	☐ Yes ☐ No ☐ Unknown	
Conjunctivitis	☐ Yes ☐ No ☐ Unknown	
Muscle aches	☐ Yes ☐ No ☐ Unknown	
Joint ache	☐ Yes ☐ No ☐ Unknown	
Loss of appetite	☐ Yes ☐ No ☐ Unknown	
Nose bleed	☐ Yes ☐ No ☐ Unknown	
Fatigue	☐ Yes ☐ No ☐ Unknown	
Chest pain	☐Yes ☐No ☐Unknown	
5d. Neurological symptoms		
Seizures	☐Yes ☐No ☐Unknown	
Altered level of consciousness	☐Yes ☐No ☐Unknown	
Other neurological signs	☐Yes ☐No ☐Unknown	
	If Yes, specify:	
5e. Other symptoms		
Other symptoms	☐ Yes ☐ No ☐ Unknown	
other symptoms	If Yes, specify:	

6. Primary health-care center/treating physician's details		
Date of first primary health facility visit (including traditional care) (dd/mm/yyyy)	// ☐ Not applicable (NA) ☐ Unknown	
Total health facilities visited to date	□ NA □ Unknown	
Total Health facilities visited to date	Specify:	
Visit to primary healthcare (PHC; GP, etc) (repeat for as many visits as required)	☐Yes ☐No ☐Unknown	
If yes, date of first PHC contact (dd/mm/yyyy)	// UnknownNA	
If yes, Name of health-care center		
If yes, Name of treating physician		
If yes, Telephone number		
If yes, Fax		
If yes, Address		
7. Other health-care interactions		
Contact with emergency number/ hotline	☐Yes ☐No ☐Unknown	
Date of emergency contact (dd/mm/yyyy)	/	
Visited emergency department (A&E) (repeat for as many contacts as required)	☐ Yes ☐ No ☐ Unknown	
Date of first A&E contact (dd/mm/yyyy)	//   Unknown	
Hospitalization (repeat for as many admissions as required)	☐Yes ☐No ☐Unknown	
Date of admission to hospital (dd/mm/yyyy)	//   Unknown	
Hospitalization (repeat for as many admissions as required)	☐Yes ☐No ☐Unknown	
Date of admission to hospital (dd/mm/yyyy)	/NA	
Name of hospital		
Location of hospital		
Date of discharge from hospital (dd/mm/yyyy)	/	
Reason for hospitalization	☐ Isolation/quarantine ☐ Clinical management ☐ Other If Other, specify:	
ICU (intensive care unit) admission	☐Yes ☐No ☐Unknown	
Date of ICU admission (dd/mm/yyyy)	// UnknownNA	
Date of discharge from ICU (dd/mm/yyyy)	/or Ongoing Unknown NA	
Mechanical ventilation	☐Yes ☐No ☐Unknown	
Dates of mechanical ventilation (dd/mm/yyyy)	Start:/ or Ongoing  Unknown NA	
Length of ventilation (days)		

8. Case symptoms: complications	
Acute respiratory distress syndrome (ARDS)	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://
Acute renal failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://
Cardiac failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://
Consumptive coagulopathy	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://
Pneumonia by chest X-ray	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://
Other complications	☐ Yes ☐ No ☐ Unknown
	If Yes, secondary bacterial infection*  ☐ Yes ☐ No ☐ Unknown  Specify infection:
	If other complication, specify:
	* Fill out relevant laboratory information in specimen collection form
Hypotension requiring vasopressors	☐ Yes ☐ No ☐ Unknown
Extracorporeal membrane oxygenation (ECMO) required	☐ Yes ☐ No ☐ Unknown
Outcome (current)	☐ Alive☐ Dead, if Yes, specify date of death (dd/mm/yyyy):☐ ☐ Dead, if Yes, specify date of death (dd/mm/yyyy):☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐
	Unknown/lost to follow-up
9. Treatment with antivirals	
Did the case receive an antiviral treatment in the last 14 days?	☐ Yes ☐ No ☐ Unknown
	(If no or unknown skip to next question)
	If Yes Specify which antiviral was received?
	Date started® (dd/mm/yyyy):// Date stopped® (dd/mm/yyyy)://
	Dosage (specify):
	What were antivirals prescribed for?  ☐ Treatment ☐ Prophylaxis

10. Patient pre-existing condition(s)	
Pregnancy	☐ Yes ☐ No ☐ Unknown If Yes, specify trimester: ☐ First ☐ Second ☐ Third ☐ Unknown
Obesity	☐Yes ☐No ☐Unknown
Cancer	☐Yes ☐No ☐Unknown
Diabetes	☐ Yes ☐ No ☐ Unknown
HIV/other immune deficiency	☐ Yes ☐ No ☐ Unknown
Heart disease	☐ Yes ☐ No ☐ Unknown
Asthma (requiring medication)	☐ Yes ☐ No ☐ Unknown
Chronic lung disease (non-asthma)	☐ Yes ☐ No ☐ Unknown
Chronic liver disease	☐ Yes ☐ No ☐ Unknown
Chronic haematological disorder	☐ Yes ☐ No ☐ Unknown
Chronic kidney disease	☐ Yes ☐ No ☐ Unknown
Chronic neurological impairment/disease	☐ Yes ☐ No ☐ Unknown
Organ or bone marrow recipient	☐ Yes ☐ No ☐ Unknown
Other pre-existing condition(s)	☐ Yes ☐ No ☐ Unknown If Yes, specify:
11. Vaccination	
Case has been vaccinated for pathogen X/disease X in the 12 months prior to onset of symptoms  Comment: this field will need to be adapted to include a vaccine for pathogen/disease X (if one is available), or to include others that may be associated with pathogen/disease X	☐ Yes ☐ No ☐ Unknown  If Yes, date of vaccination, country of vaccination (dd/mm/yyyy)://  Country:
Case was vaccinated for seasonal influenza in the 12 months prior to onset of symptoms	☐ Yes ☐ No ☐ Unknown  If Yes, date of vaccination, country of vaccination  (dd/mm/yyyy)://  Country:
Case was vaccinated for SARS-CoV-2 in the 12 months prior to onset of symptoms	☐ Yes ☐ No ☐ Unknown  If Yes, date of vaccination, country of vaccination (dd/mm/yyyy):/  Country:  Date of last infection (dd/mm/yyyy):/ or ☐ Unknown
Case was vaccinated with pneumococcal vaccine	☐ Yes ☐ No ☐ Unknown  If Yes, date (dd/mm/yyyy)  ——/——/

12. Human exposures in the days before symptom onset (in the past 14 days)		
Case travelled within the last 14 days domestically?	☐ Yes ☐ No ☐ Unknown If Yes, dates of travel (dd/mm/yyyy):/to/  Regions visited:  Cities visited:	
Case travelled within the last 14 days internationally?	☐ Yes ☐ No ☐ Unknown If Yes, dates of travel (dd/mm/yyyy): / to/  Regions visited:  Cities visited:	
Has the case had contact with anyone with suspected or confirmed pathogen X infection in the past 10 days?	☐ Yes ☐ No ☐ Unknown If Yes, dates of last contact (dd/mm/yyyy)://	
If yes, location of exposure	☐ Home ☐ Hospital ☐ Workplace ☐ Tour group ☐ School ☐ Unknown ☐ Other, specify:	
Has the case attended a festival or mass gathering in the past 14 days?	☐ Yes ☐ No ☐ Unknown If Yes, specify:	
Has the case visited or been admitted to an inpatient health facility in the past 14 days?	☐ Yes ☐ No ☐ Unknown If Yes, specify:	
Has the case visited an outpatient treatment facility in the past 14 days?	☐ Yes ☐ No ☐ Unknown If Yes, specify:	
Has the case visited a traditional healer in the past 14 days?	☐ Yes ☐ No ☐ Unknown If Yes, specify type:	
13. Status of form completion		
Form completed		

# **OPTIONAL INFORMATION TO COLLECT**

Please note there is no need to collect this information if the pathogen is known to sustainably transmit between humans. Only collect this information if there is suspected ongoing zoonotic transmission. Note: cases with zoonotic exposure can't be included in the same analysis of transmission parameters as human acquired cases.

14. Human exposures to animats in the days before fitness offset (in the past 14 days)			
Direct handling of animals			
A	Case handled animals	☐ Yes ☐ No ☐ Unknown If No or Unknown, skip to F If Yes, specify:	
В	Types of animals handled	☐ Pigs ☐ Chicken ☐ Ducks ☐ Other If other, specify:	
С	Nature of contact (e.g., feed, groom or slaughter)	Specify:	
D	Location of animal contact (select all that are relevant)	☐ Home ☐ Workplace ☐ Neighbourhood ☐ Live animal market ☐ Agricultural fair/zoo group ☐ Farm ☐ Other, specify:	
E	Within 2 weeks before or after this contact, were any animals sick or dead in the location of animal contact?	☐ Yes ☐ No ☐ Unknown If Yes, specify type and number, and proportion from flock or herd:	
Oth	er contact (not direct handling)		
F	Case had no direct contact with animals (e.g., in neighborhood, farm, zoo, at home, agricultural fair or work)	☐ Yes ☐ No ☐ Unknown If No or Unknown, skip to J If Yes, specify:	
G	Types of animals in that environment	☐ Pigs ☐ Chicken ☐ Ducks ☐ Other If other, specify:	
Н	Location of exposure	☐ Home ☐ Workplace ☐ Neighbourhood ☐ Live animal market ☐ Agricultural fair/zoo group ☐ Farm ☐ Other, specify:	
I	Within 2 weeks before or after exposure, were any animals sick or dead in the location of possible exposure?	☐ Yes ☐ No ☐ Unknown If Yes, specify type and number, and proportion from flock or herd:	
J	Case exposed to animal by-products (e.g., bird feathers) or animal excreta	☐ Yes ☐ No ☐ Unknown If Yes, specify:	

# The First Few X cases and contacts (FFX) investigation template protocol for respiratory pathogens with pandemic potential



Form A2: Case follow-up reporting form – for pathogen X index cases (Day 28)

Unique Case ID/Cluster number (if applicable):	
1. Data collector information	
Name of data collector	
Data collector institution	
Data collector telephone number	
Data collector email	
Form completion date (dd/mm/yyyy)	
2. Interview respondent information (if the person	providing the information is not the index case)
First name	
Family name	
Sex	☐ Male ☐ Female ☐ Non-binary ☐ Not known
Date of birth (dd/mm/yyyy)	/ Unknown
Relationship to case (select all that are relevant)	☐ Household member   ☐ Immediate family member   ☐ Extended family member   ☐ Health-care worker looking after case   ☐ Friend   ☐ Co-worker   ☐ Teacher   ☐ Carer   ☐ Acquaintance   ☐ Unknown
Respondent address	
Telephone (mobile) number	
3. Outcome/status	
Status	☐ Alive☐ Dead, if Yes, specify date of death (dd/mm/yyyy):☐ ☐ Unknown/lost to follow-up
Hospitalization ever required?	☐ Yes ☐ No ☐ Unknown

## Form A2: Case follow-up reporting form – for pathogen X index cases (Day 28) (continued)

(NB: If the information below is not currently available, please leave blank and send through an update as soon as results are available)		
If dead, contribution of Pathogen X to death:	☐ Underlying/primary ☐ Contributing/secondary ☐ No contribution to death ☐ Unknown	
If dead, was a post mortem performed?	☐ Yes ☐ No ☐ Unknown	
If dead, results of postmortem's report where available:		
If dead, cause of death on Death certificate (specify)		
4. Hospital health-care interactions since baseline		
Hospitalization since baseline	☐ Yes ☐ No ☐ Unknown	
Date of admission to hospital (dd/mm/yyyy)	// Unknown	
Name of hospital		
Location of hospital		
Date of discharge from hospital (dd/mm/yyyy)	/or 🗌 Ongoing	
Reason for hospitalization	☐ Isolation/quarantine ☐ Clinical management ☐ Other If Other, specify:	
ICU (intensive care unit) admission	☐Yes ☐No ☐Unknown	
Date of ICU admission (dd/mm/yyyy)	// Unknown	
Date of discharge from ICU (dd/mm/yyyy)	/or □ Ongoing □ Unknown □ NA	
Mechanical ventilation since baseline	☐Yes ☐No ☐Unknown	
Dates of mechanical ventilation (dd/mm/yyyy)	Start:/ or Ongoing  Unknown NA	
Length of ventilation (days)		
5. Case symptoms: complications		
Acute respiratory distress syndrome (ARDS)	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://	
Acute renal failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://	
Cardiac failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://	
Consumptive coagulopathy	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://	
Pneumonia by chest X-ray	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)://	

#### Form A2: Case follow-up reporting form – for pathogen X index cases (Day 28) (continued)

Other complications	☐Yes ☐ No ☐ Unknown
	If Van and and have been sided in faction *
	If Yes, secondary bacterial infection*  ☐ Yes ☐ No ☐ Unknown
	Specify infection:
	Specify infection.
	If other complication, specify:
	* Fill out relevant laboratory information in specimen collection form
Hypotension requiring vasopressors	☐ Yes ☐ No ☐ Unknown
Extracorporeal membrane oxygenation (ECMO) required	☐ Yes ☐ No ☐ Unknown
6. Patient pre-existing condition(s)	
Pregnancy	☐Yes ☐ No ☐ Unknown
	If Yes, specify trimester:
	☐ First ☐ Second ☐ Third ☐ Unknown
7. Treatment with antivirals	
Did the case receive an antiviral treatment in the last	☐ Yes ☐ No ☐ Unknown
14 days (or since baseline)?	(If No or Unknown, skip to next question)
	If Van Juliah anticipal Julian parasita da
	If Yes, which antiviral was received?
	Date started (dd/mm/yyyy)//
	Date stopped (dd/mm/yyyy)//
	Dosage (specify):
	What was anti-independent of fact
	What were antivirals prescribed for?  ☐ Treatment ☐ Prophylaxis
8. Final case classification	
Final case classification	☐ Confirmed ☐ Probable ☐ Suspected
	&
	☐ Primary ☐ Co-primary ☐ Secondary ☐ Other, specify: (e.g., tertiary case)
	& (optional)
	☐Imported
	& (optional)
	□Unrelated

## Form A2: Case follow-up reporting form – for pathogen X index cases (Day 28) (continued)

9. Status of form completion	
Form completed	☐ Yes ☐ No or partially
	If No or partially, reason:  ☐ Missed ☐ Not attempted ☐ Not performed ☐ Refusal ☐ Other, specify:

The First Few X cases and contacts (FFX) investigation template protocol for respiratory pathogens with pandemic potential

#### 2. For contacts



Unique Index Case ID/Cluster number (if applicable):	
Contact ID Number (C):  Note: Contact ID numbers should be issued at the time of completion of Form A1/Contact Line List.	
1. Current status	
☐ Alive ☐ Dead ☐ Unknown/lost to follow-up	
2. Data collector information	
Name of data collector	
Data collector institution	
Data collector telephone number	
Data collector email	
Form completion date (dd/mm/yyyy)	
3. Interview respondent information (if the person	providing the information is not the contact)
First name	
Family name	
Sex	☐ Male ☐ Female ☐ Non-binary ☐ Not known
Date of birth (dd/mm/yyyy)	/
Relationship to contact (select all that are relevant)	☐ Household member
	☐ Immediate family member
	Extended family member
	☐ Health-care worker looking after contact☐ Friend
	Co-worker
	☐ Teacher
	 ☐ Carer
	☐Acquaintance
	□Unknown
Respondent address	
Telephone (mobile) number	

4. Contact details (details of the contact)	
First name*	
Family name*	
Sex	☐ Male ☐ Female ☐ Non-binary ☐ Not known
Date of birth (dd/mm/yyyy)*	/
Relationship to case (select all that are relevant)	☐ Household member ☐ Immediate family member ☐ Extended family member ☐ Health-care worker looking after contact ☐ Friend ☐ Co-worker ☐ Teacher ☐ Carer ☐ Acquaintance ☐ Unknown
Address (village/town, district, province/region)*	
Telephone (mobile) number	
Email	
Preferred mode of contact	☐ Mobile ☐ Work ☐ Home ☐ Email
Nationality	
Country of residence	
National social number/identifier (optional)*	
Ethnicity (optional)	☐ Arab ☐ Black ☐ East Asian ☐ South Asian ☐ West Asian ☐ Latin American ☐ White ☐ Aboriginal/First Nations ☐ Other: ☐ Unknown
<ul> <li>* These identifiers are commonly accepted as personally identifiable info and should be updated by Country Y according to national guidelines.</li> <li>5. General exposure information</li> </ul>	rmation and must be kept confidential, however these may vary by country
Have you travelled within the last 14 days domestically?	☐ Yes ☐ No ☐ Unknown If Yes, dates of travel (dd/mm/yyyy): / to/ Regions visited:  Cities visited:
Have you travelled within the last 14 days internationally?	☐ Yes ☐ No ☐ Unknown If Yes, dates of travel (dd/mm/yyyy): / to/ Countries visited:  Cities visited:

In the past 14 days, have you had contact with any suspected or confirmed pathogen X cases (i.e., other cases that are not the index case in their recruited cluster)	☐ Yes ☐ No ☐ Unknown If Yes, dates of last contact (dd/mm/yyyy)://
Occupation (specify location/facility)	☐ Health worker ☐ Working with animals ☐ Health laboratory worker ☐ Other, specify:  For each occupation, please specify location or facility:

#### Note for next 2 sections:

- Complete Section 6 if the contact is a health worker (HW).
- Complete Section 7 if the contact is NOT a health worker.

6. Exposure information (if the close contact is a Health Worker (HW))		
Job title (specify)		
Place of work		
Direct physical contact with the index case (e.g., hands-on physical contact)	☐Yes ☐No	
Has the HW had prolonged face-to-face contact (>15 minutes) with a case in a health facility?	☐ Yes ☐ No  If Yes, what type of protective equipment was used by the HW? (select all that are relevant)	
	☐ Gown ☐ Gloves ☐ Eye protection ☐ Surgical/medical mask ☐ NIOSH-certified N95 or an EU standard FFP2 mask ☐ FFP3 mask	
Was the contact present while any aerosol-generating procedures took place?	☐ Yes ☐ No If Yes, specify procedure and date (dd/mm/yyyy)	
(Add as many procedures and their dates as required)	Procedure://  Was the contact wearing any type of a mask at this/these procedures? (select all that are relevant)  Surgical/medical mask  NIOSH-certified N95, an EU standard FFP2 mask  FFP3 mask	

7. Exposure information (if the close contact is NOT	a Health Worker)	
Is the contact a household contact?	☐ Yes ☐ No If Yes, specify household ID/ number:	
Specify characteristics of contact with the case from first contact, while the case was <b>symptomatic</b> , until the last unprotected contact	Date (dd/mm/yyyy)	// Still in contact with case
unprotected contact	Duration	(minutes)
(Add as many dates as required)	Setting	☐ Home/household ☐ Hospital/health care ☐ Workplace ☐ Tour group ☐ Other, specify:
8a. Symptoms in contact		
Has the contact experienced any respiratory symptoms (sore throat, runny nose, cough, shortness of breath) in the period from 10 days <b>before</b> symptom onset in the index case until the present?	☐ Yes ☐ No	
Has the contact experienced any respiratory symptoms (sore throat, runny nose, cough, shortness of breath) in the period up to 10 days <b>after</b> the last contact or until the present date, whichever is the earlier?	☐ Yes ☐ No	
Date (dd/mm/yyyy) and time of first symptom onset	//	m
Fever (>38 °C) or history of fever since disease onset	☐ Yes ☐ No ☐ Unknown If Yes, date//	
8b. Respiratory symptoms		
Sore throat	☐ Yes ☐ No ☐ Unknown If Yes, date//	
Runny nose	☐ Yes ☐ No ☐ Unknown If Yes, date//	
Cough	☐ Yes ☐ No ☐ Unknown If Yes to cough, productive, ☐ If Yes to cough, dry, ☐ Yes ☐  If Yes, date//	
Shortness of breath	☐ Yes ☐ No ☐ Unknown If Yes, date//	

8c. General symptoms	
Chills	☐Yes ☐No ☐Unknown
Anosmia (loss of smell)	☐Yes ☐No ☐Unknown
Ageusia (loss of taste)	☐Yes ☐No ☐Unknown
Vomiting	☐Yes ☐No ☐Unknown
Nausea	☐Yes ☐No ☐Unknown
Diarrhoea	☐Yes ☐No ☐Unknown
Headache	☐Yes ☐No ☐Unknown
Rash	☐Yes ☐No ☐Unknown
Conjunctivitis	☐Yes ☐No ☐Unknown
Muscle aches	☐Yes ☐No ☐Unknown
Joint ache	☐Yes ☐No ☐Unknown
Loss of appetite	☐Yes ☐No ☐Unknown
Nose bleed	☐Yes ☐No ☐Unknown
Fatigue	☐Yes ☐No ☐Unknown
Chest pain	☐Yes ☐No ☐Unknown
8d. Neurological symptoms	
Seizures	☐Yes ☐No ☐Unknown
Altered level of consciousness	☐Yes ☐No ☐Unknown
Other neurological signs	☐ Yes ☐ No ☐ Unknown If Yes, specify:
8e. Other symptoms	
Other symptoms	☐ Yes ☐ No ☐ Unknown If Yes, specify:
9. If ill, health-care center/treating physician's deta	ails
Date of first health facility visit (including traditional care) (dd/mm/yyyy)	/
Total health facilities visited to date	□ NA □ Unknown Specify:
Visit to primary healthcare (PHC; GP, etc) (repeat for as many visits as required)	☐ Yes ☐ No ☐ Unknown
If yes, date of first PHC contact (dd/mm/yyyy)	Unknown NA
If yes, Name of health-care center	
If yes, Name of treating physician	
If yes, Telephone number	
If yes, Fax	
If yes, Address	

10. If ill, other health-care interactions	
Contact with emergency number/ hotline	☐Yes ☐ No ☐ Unknown
Date of emergency contact (dd/mm/yyyy)	/
Visited emergency department (A&E) (repeat for as many contacts as required)	☐ Yes ☐ No ☐ Unknown
Date of first A&E contact (dd/mm/yyyy)	// UnknownNA
Hospitalization (repeat for as many admissions as required)	☐ Yes ☐ No ☐ Unknown
Date of admission to hospital (dd/mm/yyyy)	// UnknownNA
Name and place of hospital	
Date of discharge from hospital (dd/mm/yyyy)	/ or □Ongoing □Unknown □NA
Reason for hospitalization	☐ Isolation/quarantine ☐ Clinical management ☐ Other If Other, specify:
ICU (intensive care unit) admission	☐ Yes ☐ No ☐ Unknown
Date of ICU admission (dd/mm/yyyy)	// Unknown
Date of discharge from ICU (dd/mm/yyyy)	/ or □Ongoing □Unknown □NA
Mechanical ventilation	☐ Yes ☐ No ☐ Unknown
Dates of mechanical ventilation (dd/mm/yyyy)	Start://         Stop:// or □ Ongoing         □ Unknown
Length of ventilation (days)	
11. If ill, contact symptoms: complications	
Acute respiratory distress syndrome (ARDS)	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Acute renal failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Cardiac failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Consumptive coagulopathy	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Pneumonia by chest X-ray	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Other complications	☐ Yes ☐ No ☐ Unknown  If Yes, secondary bacterial infection* ☐ Yes ☐ No ☐ Unknown  Specify infection:  If other complication, specify:  *Fill out relevant laboratory information in specimen collection form

Hypotension requiring vasopressors   Yes   No   Unknown    Extracorporeal membrane oxygenation (ECMO) required   Yes   No   Unknown    Outcome   Alive   Dead, if Yes, specify date of death (dd/mm/yyyy):		
Outcome     Alive     Dead, if Yes, specify date of death (dd/mm/yyyy): /   Unknown/lost to follow-up    Outcome current as of date (dd/mm/yyyy)   /   Unknown   NA    12. Contact pre-existing condition(s)   Yes   No   Unknown   If Yes, specify trimester:   First   Second   Third   Unknown	Hypotension requiring vasopressors	☐Yes ☐No ☐Unknown
Dead, if Yes, specify date of death (dd/mm/yyyy):	Extracorporeal membrane oxygenation (ECMO) required	☐Yes ☐No ☐Unknown
Unknown   NA	Outcome	Dead, if Yes, specify date of death (dd/mm/yyyy):
Pregnancy   Yes	Outcome current as of date (dd/mm/yyyy)	/ UnknownNA
Pregnancy   Yes		
If Yes, specify trimester:    First   Second   Third   Unknown	12. Contact pre-existing condition(s)	
First   Second   Third   Unknown	Pregnancy	☐Yes ☐No ☐Unknown
Obesity		
Cancer		☐ First ☐ Second ☐ Third ☐ Unknown
Diabetes	Obesity	☐ Yes ☐ No ☐ Unknown
HIV/other immune deficiency	Cancer	☐ Yes ☐ No ☐ Unknown
Heart disease	Diabetes	☐ Yes ☐ No ☐ Unknown
Asthma (requiring medication)  Chronic lung disease (non-asthma)  Chronic liver disease  Chronic haematological disorder  Chronic kidney disease  Chronic neurological impairment/disease  Organ or bone marrow recipient  Other pre-existing condition(s)  Pes No Unknown  Unknown  Unknown  Organ Unknown  Organ	HIV/other immune deficiency	☐ Yes ☐ No ☐ Unknown
Chronic lung disease (non-asthma)  Chronic liver disease  Chronic haematological disorder  Chronic kidney disease  Chronic neurological impairment/disease  Organ or bone marrow recipient  Other pre-existing condition(s)  Chronic lunknown  Wes No Unknown  Orea No Unknown  Orea No Unknown  Other pre-existing condition(s)	Heart disease	☐ Yes ☐ No ☐ Unknown
Chronic liver disease	Asthma (requiring medication)	☐ Yes ☐ No ☐ Unknown
Chronic haematological disorder	Chronic lung disease (non-asthma)	☐Yes ☐No ☐Unknown
Chronic kidney disease	Chronic liver disease	☐Yes ☐No ☐Unknown
Chronic neurological impairment/disease       ☐ Yes ☐ No ☐ Unknown         Organ or bone marrow recipient       ☐ Yes ☐ No ☐ Unknown         Other pre-existing condition(s)       ☐ Yes ☐ No ☐ Unknown	Chronic haematological disorder	☐Yes ☐No ☐Unknown
Organ or bone marrow recipient	Chronic kidney disease	☐Yes ☐No ☐Unknown
Other pre-existing condition(s)	Chronic neurological impairment/disease	☐Yes ☐No ☐Unknown
	Organ or bone marrow recipient	☐Yes ☐No ☐Unknown
	Other pre-existing condition(s)	

13. Vaccination	
Contact has been vaccinated for pathogen X/disease X in the 12 months prior to onset of symptoms  Comment: this field will need to be adapted to include a vaccine for pathogen/disease X (if one is available), or to include others that may be associated with pathogen/disease X	☐ Yes ☐ No ☐ Unknown  If Yes, date of vaccination, country of vaccination (dd/mm/yyyy)://  Country:
Contact was vaccinated for seasonal influenza in the 12 months prior to onset of symptoms	☐ Yes ☐ No ☐ Unknown  If Yes, date of vaccination, country of vaccination  (dd/mm/yyyy)://  Country:
Contact was vaccinated for SARS-CoV-2 in the 12 months prior to onset of symptoms	☐ Yes ☐ No ☐ Unknown  If Yes, date of vaccination, country of vaccination (dd/mm/yyyy)://  Country:  Date of last infection (dd/mm/yyyy):/ or ☐ Unknown
Contact was vaccinated with pneumococcal vaccine	☐ Yes ☐ No ☐ Unknown If Yes, date (dd/mm/yyyy)//
14. Status of form completion	
Form completed	☐ Yes ☐ No or partially  If No or partially, reason: ☐ Missed ☐ Not attempted ☐ Not performed ☐ Refusal ☐ Other, specify:

# The First Few X cases and contacts (FFX) investigation template protocol for respiratory pathogens with pandemic potential



**Form B2:** Contact follow-up reporting form – for close contacts of pathogen X index cases (Day 28)

Unique Index Case ID/Cluster number (if applicable):	
Contact ID Number (C):  Note: Contact ID numbers should be issued at the time of completion of Form A1/Contact Line List.	
1. Data collector information	
Name of data collector	
Data collector institution	
Data collector telephone number	
Data collector email	
Form completion date (dd/mm/yyyy)	/
2. Interview respondent information (if the persor	providing the information is not the contact)
First name	
Family name	
Sex	☐ Male ☐ Female ☐ Non-binary ☐ Not known
Date of birth (dd/mm/yyyy)	/
Relationship to contact (select all that are relevant)	☐ Household member
	☐ Immediate family member
	Extended family member
	☐ Health-care worker looking after contact
	Friend
	☐ Co-worker
	Teacher
	Carer
	☐ Acquaintance ☐ Unknown
P de de delle de	
Respondent address	
Talanhana (mahila) numbar	
Telephone (mobile) number	

3. Exposure information (update from baseline)					
Specify characteristics of contact with the confirmed	Date (dd/mm/yyyy)	//			
case from first contact, while the primary case was <b>symptomatic</b> , until the last unprotected contact	Duration	(minutes)  Still in contact with case			
(Add as many dates as required)	Setting	☐ Home/household (Household ID number:) ☐ Hospital/health care ☐ Workplace ☐ Tour group ☐ Other, specify:			
4. Outcome/status					
Status	☐ Alive☐ Dead, if Yes, specify date of ☐ Dead if Yes, specify date of ☐ Unknown/lost to follow-up				
Hospitalization ever required?	☐ Yes ☐ No ☐ Unknown				
(NB. If the information below is not currently available, presults are available)	blease leave blank and send th	nrough an update as soon as			
If dead, contribution of Pathogen X to death:	☐ Underlying/primary ☐ Contributing/secondary ☐ No contribution to death ☐ Unknown				
If dead, was a post mortem performed?	☐ Yes ☐ No ☐ Unknown				
If dead, results of postmortem's report where available:					
If dead, cause of death on Death certificate (specify)					
5. Hospital health-care interactions since baseline					
Hospitalization since baseline	☐Yes ☐No ☐Unknown				
Date of first hospitalization (dd/mm/yyyy)	// UnknownNA				
Name and place of hospital					
Date of discharge from hospital (dd/mm/yyyy)	/ or ☐ Ongo	ing			
Reason for hospitalization	☐ Isolation/quarantine ☐ Clinical management ☐ Other If Other, specify:				
ICU (intensive care unit) admission since baseline	☐ Yes ☐ No ☐ Unknown				
Date of ICU admission (dd/mm/yyyy)	// Unknown				
Date of discharge from ICU (dd/mm/yyyy)	/ or □Ongo □Unknown □NA	ing			

Mechanical ventilation since baseline	☐ Yes ☐ No ☐ Unknown
Dates of mechanical ventilation (dd/mm/yyyy)	Start://         Stop:// or □ Ongoing         □ Unknown
Length of ventilation (days)	
6. Contact symptoms: complications	
Acute respiratory distress syndrome (ARDS)	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Acute renal failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Cardiac failure	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Consumptive coagulopathy	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Pneumonia by chest X-ray	☐ Yes ☐ No ☐ Unknown If Yes, date started (dd/mm/yyyy)//
Other complications	☐ Yes ☐ No ☐ Unknown  If Yes, secondary bacterial infection* ☐ Yes ☐ No ☐ Unknown  Specify infection:
	If other complication, specify:
Hypotension requiring vasopressors	*Fill out relevant laboratory information in specimen collection form  Yes No Unknown
Extracorporeal membrane oxygenation (ECMO) required	☐ Yes ☐ No ☐ Unknown
- · · · · · · · · · · · · · · · · · · ·	
7. Contact pre-existing condition(s)	
Pregnancy	☐ Yes ☐ No ☐ Unknown  If Yes, specify trimester: ☐ First ☐ Second ☐ Third ☐ Unknown
8. Treatment with antivirals	
Did the contact receive an antiviral treatment in the last 14 days (or since baseline)?	☐ Yes ☐ No ☐ Unknown  (If No or Unknown, skip to next question)
	If Yes, which antiviral was received?
	Date started (dd/mm/yyyy)// Date stopped (dd/mm/yyyy)//
	Dosage (specify):
	What were antivirals prescribed for?  ☐ Treatment ☐ Prophylaxis

9. Final contact classification	
Contact was identified as a case during the investigation	☐ Yes ☐ No  If no, the contact is a non-case
Final case classification	Confirmed Probable Suspected  & Primary Co-primary Secondary Other, specify: (e.g., tertiary case)  & (optional) Imported  & (optional) Unrelated
10. Status of form completion	
Form completed	☐ Yes ☐ No or partially  If No or partially, reason: ☐ Missed ☐ Not attempted ☐ Not performed ☐ Refusal ☐ Other, specify:

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#### 3. For cases and contacts



Form C: Specimen collection forms and laboratory results

**Comment:** Please note that this table will need to be filled/ updated for each respiratory and serological specimen collected per the mandatory sampling strategy

1. Case and contact details	
Unique ID for participant (use unique case or contact ID)	
Classification at recruitment	☐ Index case ☐ Co-index case ☐ Close contact
2. Respiratory specimen collection	
Date of sample collection (dd/mm/yyyy)	/
Day of specimen collection per sampling strategy	(insert day number)
Type of sample collection	☐ Nasal swab ☐ Throat swab ☐ Nasopharyngeal swab ☐ Other, specify
Who collected the respiratory specimen?	☐ Study staff/ research nurse ☐ Self-collected ☐ Other professional specimen collection service
Which laboratory was the specimen sent to?	
Date sample received by laboratory (dd/mm/yyyy)	/
Laboratory identification number	
Diagnostic method	☐ PCR ☐ Viral culture or virus isolation ☐ Other, specify
Date of result	
Pathogen X Result	☐ Positive ☐ Negative ☐ Inconclusive/probable  If positive (and applicable), subtype* ☐ Subtype A ☐ Subtype B ☐ Subtype C ☐ Not able to be typed  *Adapt based on known information on pathogen X

## Form C: Specimen collection forms and laboratory results (continued)

Other laboratory results	☐ Other pathogens, specify#:
	☐ Negative ☐ NA / not tested  # Include any secondary (bacterial) infections in this section
Sample sent to WHO CC (if applicable) (dd/mm/yyyy)	☐Yes
	□No
	If yes, Date:/
	Name of collaborating centre:
3. Serology testing methods and results:	
Date of sample collection (dd/mm/yyyy)	
Day of specimen collection per sampling strategy	(insert day number)
Type of sample collection	☐ Whole blood
	Serum
	☐ Dried blood spot
	Other, specify:
Who collected the sample?	☐ Study staff/ research nurse☐ Other professional specimen collection service
Which laboratory was the specimen sent to?	
Date sample received by laboratory (dd/mm/yyyy)	
Laboratory identification number	
Diagnostic method/assay used	☐ Total antibody ☐ Microneutralisation ☐ Other, specify:
Antigen used (if applicable)	
Date of result	
Pathogen X Result (according to assay thresholds/cut-offs – please report these)	☐ Positive ☐ Negative ☐ Indeterminate/probable  Titre (irrespective of result):
Sample sent to WHO CC (if applicable) (dd/mm/yyyy)	☐ Yes ☐ No
	If yes, Date:/
	Name of collaborating centre:

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**Form D:** Symptom diary for index cases of pathogen X and close contacts (Day 2–28)

Symptom diaries will be provided to all participants, for recording the presence or absence of various signs or symptoms for 28 days after the administration of the initial case and contact questionnaires (Form A1 and B1).

The symptom diary template provided below is generic.

**Comment:** 

In the context of a novel pathogen with uncertain clinical presentation and spectrum, symptom diaries may be broadened to include vomiting, diarrhoea, abdominal pain, etc., as relevant.

#### Symptom diaries

#### Case or contact ID number:

Day	Symptoms*						
	No symptoms (check if none experienced)	Fever ≥38°C	Runny nose	Cough	Sore throat	Shortness of breath	Other symptoms: specify
2	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No	
				Unknown			
3	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry?	☐ Yes ☐ No	☐ Yes ☐ No	
				☐ Yes ☐ No ☐ Unknown			

<sup>\*</sup> Please select None for No symptoms. If no symptoms are experienced, then consider the entry complete



Form D: Symptom diary for index cases of pathogen X and close contacts (Day 2–28) (continued)

Day	Symptoms*						
	No symptoms (check if none experienced)	Fever ≥38°C	Runny nose	Cough	Sore throat	Shortness of breath	Other symptoms: specify
4	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	□ Yes □ No	□ Yes □ No	
5	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	□Yes □No	□Yes □No	
6	□None	□Yes □No	□Yes □No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	□Yes □No	□Yes □No	
7	□None	□Yes □No	□Yes □No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	□ Yes □ No	□Yes □No	
8	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	□Yes □No	□ Yes □ No	

Form D: Symptom diary for index cases of pathogen X and close contacts (Day 2–28) (continued)

Day	Symptoms*						
	No symptoms (check if none experienced)	Fever ≥38°C	Runny nose	Cough	Sore throat	Shortness of breath	Other symptoms: specify
9	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	☐ Yes ☐ No	□ Yes □ No	
				Unknown			
10	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	
				If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown			
11	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	☐ Yes ☐ No	□ Yes □ No	
12	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Unknown  ☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	□ Yes □ No	
13	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	

Form D: Symptom diary for index cases of pathogen X and close contacts (Day 2–28) (continued)

Day	Symptoms*						
	No symptoms (check if none experienced)	Fever ≥38°C	Runny nose	Cough	Sore throat	Shortness of breath	Other symptoms: specify
14	□None	☐ Yes ☐ No	□Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	□ Yes □ No	□ Yes □ No	
				Unknown			
15	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	
				If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown			
16	□None	☐ Yes ☐ No	□Yes □ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	□ Yes □ No	
17	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	
18	□None	☐ Yes ☐ No	□ Yes □ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	□ Yes □ No	☐ Yes ☐ No	

Form D: Symptom diary for index cases of pathogen X and close contacts (Day 2–28) (continued)

Day	Symptoms*						
	No symptoms (check if none experienced)	Fever ≥38°C	Runny nose	Cough	Sore throat	Shortness of breath	Other symptoms: specify
19	□None	☐ Yes ☐ No	□ Yes □ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No	
				Unknown			
20	□None	☐ Yes ☐ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	
				If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown			
21	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No  If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	□ Yes □ No	
22	□None	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	
23	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	□ Yes □ No	

Form D: Symptom diary for index cases of pathogen X and close contacts (Day 2–28) (continued)

Day	Symptoms*						
	No symptoms (check if none experienced)	Fever ≥38°C	Runny nose	Cough	Sore throat	Shortness of breath	Other symptoms: specify
24	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry? ☐ Yes ☐ No	□ Yes □ No	□ Yes □ No	
				☐Unknown			
25	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown	☐ Yes ☐ No	☐ Yes ☐ No	
				If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown			
26	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown  If Yes to cough, dry?	□ Yes □ No	☐ Yes ☐ No	
				☐ Yes ☐ No ☐ Unknown			
27	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown	□Yes □No	□ Yes □ No	
				If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown			
28	□None	□ Yes □ No	□ Yes □ No	☐ Yes ☐ No If Yes to cough, productive? ☐ Yes ☐ No ☐ Unknown	□ Yes □ No	□ Yes □ No	
				If Yes to cough, dry? ☐ Yes ☐ No ☐ Unknown			

## FFX reporting forms: completion guidance

These notes provide guidance in completing the forms. It is suggested that the investigations could be divided into teams – these could include:

- a "case reporter" team;
- a "contact reporter" team; and
- a **"go to" team** who would liaise with additional data sources other than the case or contact, such as hospitals, laboratories, etc.

# Form A1: Case initial report form – for pathogen X index cases (Day 1) and Form A2: Case follow-up form – for pathogen X index cases (Day 28)

These forms should be completed by the "Case reporter" team.

Section Sources		Verified against	
Final case classification	Case reporter/hospital		
Reporter details	Case reporter		
Informant details	Informant		
Outcome/status	Informant	Statistical data, mortality, GP/hospital	
Illness	Informant	Health-care provider/review of medical records	
Clinical course/complications	Informant/interview with health-care provider	Review of medical records	
Interaction with national security system	Informant/hospital	National social health information system	
Reference test results	Testing laboratory	Laboratory database	
Bacterial infections	Testing laboratory	Laboratory database	

#### Form B1: Contact initial reporting form – for close contacts of pathogen X index cases (Day 1)

This form should be completed by the "Contacts reporter" team and should be completed after the initial case report form (B1) has been completed by the "Case reporter" team, ideally within 24 hours.

Section	Sources	Verified against
Reporter details	Contact reporter	
Informant details	Informant	
Contact details	Informant	
Exposure information	Informant	
Illness in contacts	Informant	Health-care provider/review of medical records
Outcome/status	Informant	Statistical data, mortality, GP/hospital
Case classification	Contact reporter	
Laboratory tests	Testing laboratory	Laboratory database
Medical history	Informant	Health-care provider/GP/review of medical records

#### Form B2: Contact follow-up reporting form – for close contacts of pathogen X index cases (Day28)

This form should be completed by the "Contacts reporter" team.

Section	Sources	Verified against
Reporter details	Contact reporter	
Informant details	Informant	
Final contact classification Contact reporter		
Exposure information	Informant	
Illness in contacts	Informant	Health-care provider/review of medical records
Clinical course/complications	Informant/interview with health-care provider	Review of medical records
Laboratory tests	Testing laboratory	Laboratory database

#### Form C: Specimen collection forms and laboratory results

These forms should be complete by the "go to" team.

Section Sources		Verified against	
Reference test results	Testing laboratory	Laboratory database	
Bacterial infections	Testing laboratory	Laboratory database	

#### Form D: Symptom diary for index cases of pathogen X and close contacts (Day 2-28)

This form should be completed by all participants.

## **Appendix C: Considerations for specimen collection**

Laboratory expertise should be sought for guidance on the most appropriate specimens prior to the conduct of the FFX study. Furthermore, collection and testing of appropriate specimens from cases and contacts should be conducted according to the appropriate sampling strategy.

**Table 1 (Appendix C)** provides a list of possible specimens that may be collected for identification of respiratory pathogens. The type of specimen recommended depends on the pathogen, and in some cases several specimen types may be indicated. For example, lower respiratory tract specimens are preferred for detection of MERS-CoV (30), whereas upper and lower respiratory tract specimens as well as serum are preferred for influenza virus detection (31, 32). When the event aetiology is unknown, it is useful to collect various specimens when feasible, to maximise opportunities for detection and characterisation (33).

#### **Comment:**

Investigators need to be familiar with the correct collection techniques (including the appropriate use of PPE for different types of specimens collected and established infection control guidelines), and the safety standards for specimen storage, packaging and transport.

**Appendix C Table 1: Specimen type, transportation and storage guidelines for testing for presence of respiratory pathogens.** (Adapted from Table 1, WHO protocol to investigate
non-seasonal influenza and other emerging acute respiratory diseases (33))

Specimen type	Transport medium	Transport to laboratory	Storage until testing	Comment
Nasopharyngeal wash*	N/A	4°C	≤48 hours: 4 °C >48 hours: -70 °C	
Mid-turbinate swab	VTM	4°C	≤5 days: 4 °C >5 days: -70 °C	
Nasopharyngeal swab	VTM	4°C	≤5 days: 4 °C >5 days: -70 °C	
Saliva	N/A	4°C	≤48 hours: 4 °C >48 hours: -70 °C	
Sputum	N/A	4°C	≤48 hours: 4 °C >48 hours: −70 °C	It may be difficult to ensure the material is from the lower respiratory tract
Nasal swab	VTM	4°C	≤5 days: 4 °C >5 days: -70 °C	Nasal and throat swabs may be combined to increase likelihood of detection
Throat swab	VTM	4°C	≤5 days: 4 °C >5 days: -70 °C	As above
Nasal wash*	N/A	4°C	≤48 hours: 4 °C >48 hours: -70 °C	
Nasopharyngeal aspirate*	N/A	4°C	≤48 hours: 4 °C >48 hours: –70 °C	

Specimen type	Transport medium	Transport to laboratory	Storage until testing	Comment
Bronchoalveolar lavage*	N/A	4°C	≤48 hours: 4 °C >48 hours: -70 °C	
Serum	N/A	4°C	≤5 days: 4 °C >5 days: –70 °C	Paired samples are required: acute at baseline and convalescent, 2 to 4 weeks later
Whole blood	EDTA tube	4°C	≤5 days: 4 °C >5 days: −70 °C	For antigen detection particularly during the first week of illness
Urine	N/A	4°C	≤5 days: 4 °C >5 days: –70 °C	

 $<sup>^{\</sup>star}\quad \text{More invasive specimen types which may cause patient discomfort and are not performed routinely}.$ 

N/A, Not applicable