DOLUTEGRAVIR TABLETS

DOLUTEGRAVIRI COMPRESSI

Draft proposal for inclusion in The International Pharmacopoeia

(December 2020)

DRAFT FOR COMMENTS

Please send any comments you may have on this draft working document to Dr Herbert Schmidt, Technical Officer, Norms and Standards for Pharmaceuticals, Technical Standards and Specifications (schmidt@who.int), with a copy to Ms Claire Vogel (vogelc@who.int) by 28 February 2021.

Our working documents are sent out electronically and they will also be placed on the WHO Medicines website (https://www.who.int/teams/health-product-and-policy-standards/standards-and-specifications/pharmaceuticals/current-projects) for comments under the “Working documents in public consultation” link. If you wish to receive our draft guidelines, please send your e-mail address to jonessj@who.int and your name will be added to our electronic mailing list.
SCHEDULE FOR THE PROPOSED ADOPTION PROCESS OF DOCUMENT QAS/18.780:

Draft proposal for inclusion in for *The International Pharmacopoeia*

**DOLUTEGRAVIR TABLETS**

**DOLUTEGRAVIRI COMPRESSI**

<table>
<thead>
<tr>
<th>Description of activity</th>
<th>Date</th>
</tr>
</thead>
<tbody>
<tr>
<td>First draft received from collaborating laboratory.</td>
<td>August 2018</td>
</tr>
<tr>
<td>Presentation to 53rd WHO Expert Committee on Specifications for Pharmaceutical Preparations (ECSPP).</td>
<td>October 2018</td>
</tr>
<tr>
<td>Discussion at the consultation on Screening Technologies, Laboratory Tools and Pharmacopoeial Specifications for Medicines.</td>
<td>2-3 May 2019</td>
</tr>
<tr>
<td>Draft monograph sent out for public consultation.</td>
<td>September – October 2019</td>
</tr>
<tr>
<td>Presentation to 54th WHO ECSPP.</td>
<td>October 2019</td>
</tr>
<tr>
<td>Discussion at the consultation on Screening Technologies, Laboratory Tools and Pharmacopoeial Specifications for Medicines.</td>
<td>May 2020</td>
</tr>
<tr>
<td>Draft revision 1 sent out for public consultation.</td>
<td>August – September 2020</td>
</tr>
<tr>
<td>Presentation to 55th WHO ECSPP.</td>
<td>October 2020</td>
</tr>
<tr>
<td>Draft revision 2 based on the comments received on revision 1 and the discussions at the meeting of the Expert Committee.</td>
<td>October 2020</td>
</tr>
<tr>
<td>Draft revision 2 sent out for public consultation.</td>
<td>December 2020 – February 2021</td>
</tr>
<tr>
<td>Further follow-up action as required.</td>
<td></td>
</tr>
</tbody>
</table>

[Note from the Secretariat. The monograph on dolutegravir tablets is proposed for inclusion in *The International Pharmacopoeia.*]

*Being one of the first public standard on Dolutegravir tablets, the monograph is expected to play an important role in ensuring access to quality assured dolutegravir products worldwide. Manufacturers are therefore invited to provide their feed-back on the draft monograph to help ensure that the proposed standard adequately controls the products they manufacture.*
DOLUTEGRAVIR TABLETS

DOLUTEGRAVIRI COMPRRESSI

Category. Antiretroviral (integrase strand-transfer inhibitor)

Storage. Dolutegravir tablets should be kept in a well-closed container.

Labelling. The designation of the container should state that the active ingredient is the sodium salt and the quantity should be indicated in terms of the equivalent amount of dolutegravir.


Requirements.

Comply with the monograph for Tablets.

Definition. Dolutegravir tablets contain Dolutegravir sodium. They contain not less than 90.0% and not more than 110.0% of the amount of dolutegravir (C_{20}H_{19}F_{2}N_{3}O_{5}) stated on the label.

Identity tests

- Either test A or test B may be applied.

A. Carry out the tests as specified in A. 1, or where a diode array detector is available, test A.2.

A1. Carry out the test as described under 1.14.4 High-performance liquid chromatography using the conditions and solutions given under “Assay”. The retention time of the principal peak in the chromatogram obtained with solution (1) corresponds to the retention time of the peak due to dolutegravir in the chromatogram obtained with solution (2).

To a quantity of the powdered tablets, nominally equivalent to 10 mg of dolutegravir, add 40 mL methanol R, sonicate for five minutes, allow to cool to
room temperature, dilute to 50 mL and filter. Dilute 1 mL of the filtrate to 20 mL with methanol R. The absorption spectrum (\(L_6\)) of the resulting solution, when observed between 220 nm and 400 nm, exhibits a maximum at about 258 nm.

A.2 Carry out the test as described under \textit{1.14.4 High-performance liquid chromatography} using the conditions and solutions given under “Assay”. Record the UV spectrum of the principal peak in the chromatograms with a diode array detector in the range of 220 and 400 nm. The retention time and the UV spectrum of the principal peak in the chromatogram obtained with solution (1) correspond to the retention time and the spectrum of the peak due to dolutegravir in the chromatogram obtained with solution (2).

B. Carry out the test as described under \textit{1.14.1 Thin-layer chromatography} using silica gel R6, or similar, as the coating substance and a mixture of 72 volumes of ethyl acetate R, 14 volumes of water R and 14 volumes of glacial acetic acid R as the mobile phase. Prepare as a solvent solution a mixture of 96 volumes of methanol R and 4 volumes of glacial acetic acid R. Apply separately to the plate 5 μL of each of the following two solutions. For solution (A), shake a quantity of the powdered tablets containing 10 mg of dolutegravir with 10 mL of the solvent solution and filter. For solution (B), use a solution containing 1 mg of dolutegravir sodium RS per mL solvent solution. After removing the plate from the chromatographic chamber, allow it to dry in air or in a current of cool air. Examine the chromatogram in ultraviolet light (254 nm).

The principal spot obtained with solution (A) corresponds in position, appearance and intensity with that obtained with solution (B).

After drying, spray the plate with basic potassium permanganate (5 g/L) TS. Examine the chromatogram in daylight. The principal spot obtained with solution (A) corresponds in position, appearance and intensity with that obtained with solution (B).

\textbf{Dissolution.} Carry out the test as described under \textit{5.5 Dissolution test for oral dosage forms} using as the dissolution medium 900 mL of a solution prepared by dissolving 2.5 g of sodium dodecyl sulfate R in 1000 ml dissolution buffer pH 6.8. Rotate the paddle at 50 revolutions per minute. At 45 minutes withdraw a sample of 10 mL of the medium through an in-line filter.
Allow the filtered solution to cool down to room temperature and dilute 5.0 mL of the filtered solution to 10.0 mL with dissolution medium. Use this solution as solution (1).

Measure the absorbance as described under 1.6 Spectrophotometry in the visible and ultraviolet regions in a cuvette with an optical pathlength of 10 mm layer of the resulting solutions at the maximum at about 258 nm, using the dissolution medium as the blank.

For each of the tablets tested, calculate the amount of dolutegravir (C_{20}H_{19}F_{2}N_{3}O_{5}) in the medium using the absorptivity value of 54.0 for dolutegravir sodium (A_{1\% cm} = 540). Each mg of dolutegravir sodium is equivalent to 0.950 mg of dolutegravir.

Evaluate the results as described under 5.5 Dissolution test for solid oral dosage forms, Acceptance criteria. The amount of dolutegravir in solution for each tablet is not less than 80% (Q) of the amount declared on the label.

**Related substances.** Perform the test in subdued light and without any prolonged interruptions, using low-actinic glassware.

Carry out the test as described under 1.14.4 High-performance liquid chromatography using a stainless steel column (25 cm x 4.6 mm) packed with particles of silica gel, the surface of which has been modified with chemically-bonded octadecylsilyl and pentafluorophenyl groups (5 µm).\(^1\)

Use the following conditions for gradient elution:

<table>
<thead>
<tr>
<th>Time (minutes)</th>
<th>Mobile phase A (% v/v)</th>
<th>Mobile phase B (% v/v)</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>0–2</td>
<td>60</td>
<td>40</td>
<td>Isocratic</td>
</tr>
<tr>
<td>2–32</td>
<td>60 to 50</td>
<td>40 to 50</td>
<td>Linear gradient</td>
</tr>
<tr>
<td>32–56</td>
<td>50 to 20</td>
<td>50 to 80</td>
<td>Linear gradient</td>
</tr>
</tbody>
</table>

\(^1\) An ACE 5 C18-PFP column was found suitable.
 operate at a flow rate of 1.0 mL per minute. as a detector, use an ultraviolet spectrophotometer set at a wavelength of 258 nm. maintain the column temperature at 45 °C.

prepare the following solutions using as the diluent a mixture of 60 volumes of water R and 40 volumes of acetonitrile R.

for solution (1), transfer a quantity of the powdered tablets, nominally equivalent to 70.0 mg dolutegravir, to a 100 mL volumetric flask. add about 70 mL diluent and sonicate for five minutes, cool to room temperature, dilute to volume and filter. for solution (2), dilute 1.0 mL of solution (1) to 100.0 mL. dilute 10.0 mL of this solution to 50.0 mL. for solution (3) dilute 2 mg of dolutegravir impurity D RS in 5 mL of acetonitrile R. dilute 1 mL of this solution to 10 mL. for solution (4), dissolve 5 mg of dolutegravir sodium RS in 1 mL acetonitrile R. add 4.5 mL water R and 4.5 mL hydrochloric acid (~ 420 g/L) TS and boil the solution under reflux for 1 hour. cool the solution to room temperature and dilute 1 mL of it to 10 mL with a mixture of 6 volumes of water R and 4 volumes of acetonitrile R. for solution (5) mix 1 mL of solution (3) with 1 mL of solution (4).

inject alternately 10 µL of solutions (1), (2), (3), (4) and (5).

use the chromatogram obtained with solution (3) and the chromatogram supplied with dolutegravir impurity D RS to identify the peak due to the impurity D. use the chromatogram obtained with solution (4) to identify the peak due to impurity H (the chromatogram usually shows two principal peaks: the peak due to dolutegravir and the peak due to impurity H).

the impurities, if present, are eluted at the following relative retentions with reference to dolutegravir (retention time about 30 minutes): impurity C about 0.66; impurity F about 0.70; impurity D about 0.74; impurity H about 0.78; impurity E about 0.89; impurity J about 1.75; impurity K about 1.77; impurity L about 2.10.

the test is not valid unless in the chromatogram obtained with solution (5) the resolution factor between the peaks due to impurity D and due to impurity H is at least 1.5. also, the test is not
valid unless in the chromatogram obtained with solution (2) the peak due to dolutegravir is obtained with a signal-to-noise ratio of at least 20.

In the chromatogram obtained with solution (1):

- the area of any impurity peak is not greater than the area of the peak due to dolutegravir in the chromatogram obtained with solution (2) (0.2%).
- The sum of the areas of all impurity peaks is not greater than 5 times the area of the peak due to dolutegravir in the chromatogram obtained with solution (2) (1.0%). Disregard any peak with an area less than 0.5 times the area of the peak due to dolutegravir in the chromatogram obtained with solution (2) (0.1%).

**Assay.** Perform the test in subdued light and without any prolonged interruptions, using low-actinic glassware. Carry out the test as described under 1.14.4 *High-performance liquid chromatography* using a stainless steel column (25 cm x 4.6 mm) packed with particles of silica gel, the surface of which has been modified with octadecylsilyl and perfluorophenyl groups (5 µm).²

Use the following mobile phase: Dissolve 0.186 g of disodium edetate R in 1000 mL water R and adjust to pH 3.0 with phosphoric acid (~20 g/L) TS. Mix 420 volumes of this solution with 580 volumes of methanol R.

Operate at a flow rate of 1.0 mL/minute. As a detector, use an ultraviolet spectrophotometer set at a wavelength of 258 nm. For identity test A.2, use a diode array detector in the range of 220 nm to 400 nm. Maintain the column at a temperature of 40 °C.

Prepare the following solutions using as the diluent a mixture of 60 volumes of water R and 40 volumes of acetonitrile R.

For solution (1), weigh and powder 20 tablets. Transfer a quantity of the powdered tablets, nominally equivalent to 100.0 mg of dolutegravir, to a 100 mL volumetric flask. Add about 70 mL of diluent and sonicate for five minutes, cool to room temperature and make up to volume with diluent. Filter and dilute 5.0 mL of the filtrate to 100.0 mL. For solution (2), dissolve 53.0

² An ACE 5 C18-PFP column was found suitable.
mg of dolutegravir sodium RS and dilute to 50.0 mL. Dilute 5.0 mL of this solution to 100.0 mL.

Inject alternately 20 µL each of solutions (1) and (2). Record the chromatograms for about 20 min.

Measure the areas of the peaks corresponding to dolutegravir obtained in the chromatograms of solution (1) and (2) and calculate the percentage content of dolutegravir \( \text{C}_{20}\text{H}_{18}\text{F}_{2}\text{N}_{3}\text{O}_{5} \) in the tablets using the declared content of \( \text{C}_{20}\text{H}_{18}\text{F}_{2}\text{N}_{3}\text{NaO}_{5} \) in dolutegravir sodium RS. Each mg of dolutegravir sodium is equivalent to 0.950 mg of dolutegravir.

**Impurities.** The impurities limited by the requirements of this monograph include those listed in the monograph on Dolutegravir sodium, excluding impurities A, B and G.

**International Chemical Reference Substances (ICRS) to be established:**

- **Dolutegravir sodium RS**
  - ICRS to be established.

- **Dolutegravir impurity D RS**
  - ICRS to be established.

- **Dolutegravir impurity B RS**
  - ICRS to be established.

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